



*Bogomoletz Institute of Physiology  
Kiev  
Ukraine*

Dr. Pavel Belan

***Оптогенетические методы исследования***

15.05.2014

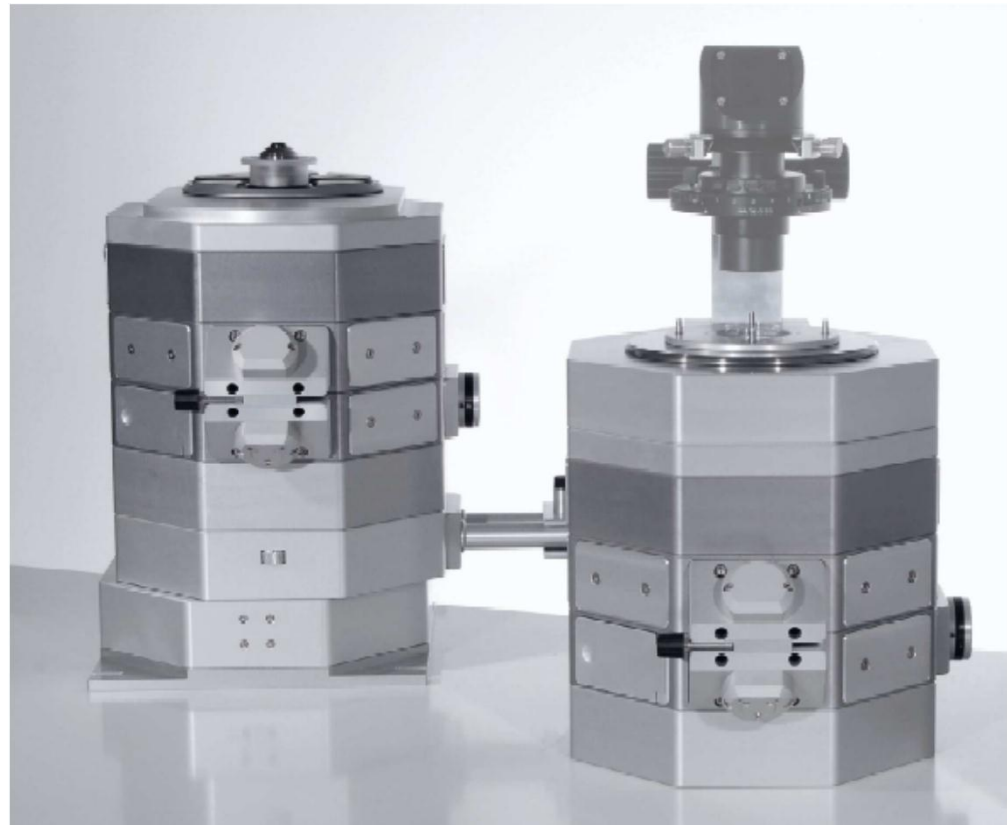
- “ *Флуоресцентная микроскопия основанная на применении флуоресцентных белков, Green Fluorescent Protein (GFP)*
- “ *Оптогенетика*

# State-of-the-Art microscopes

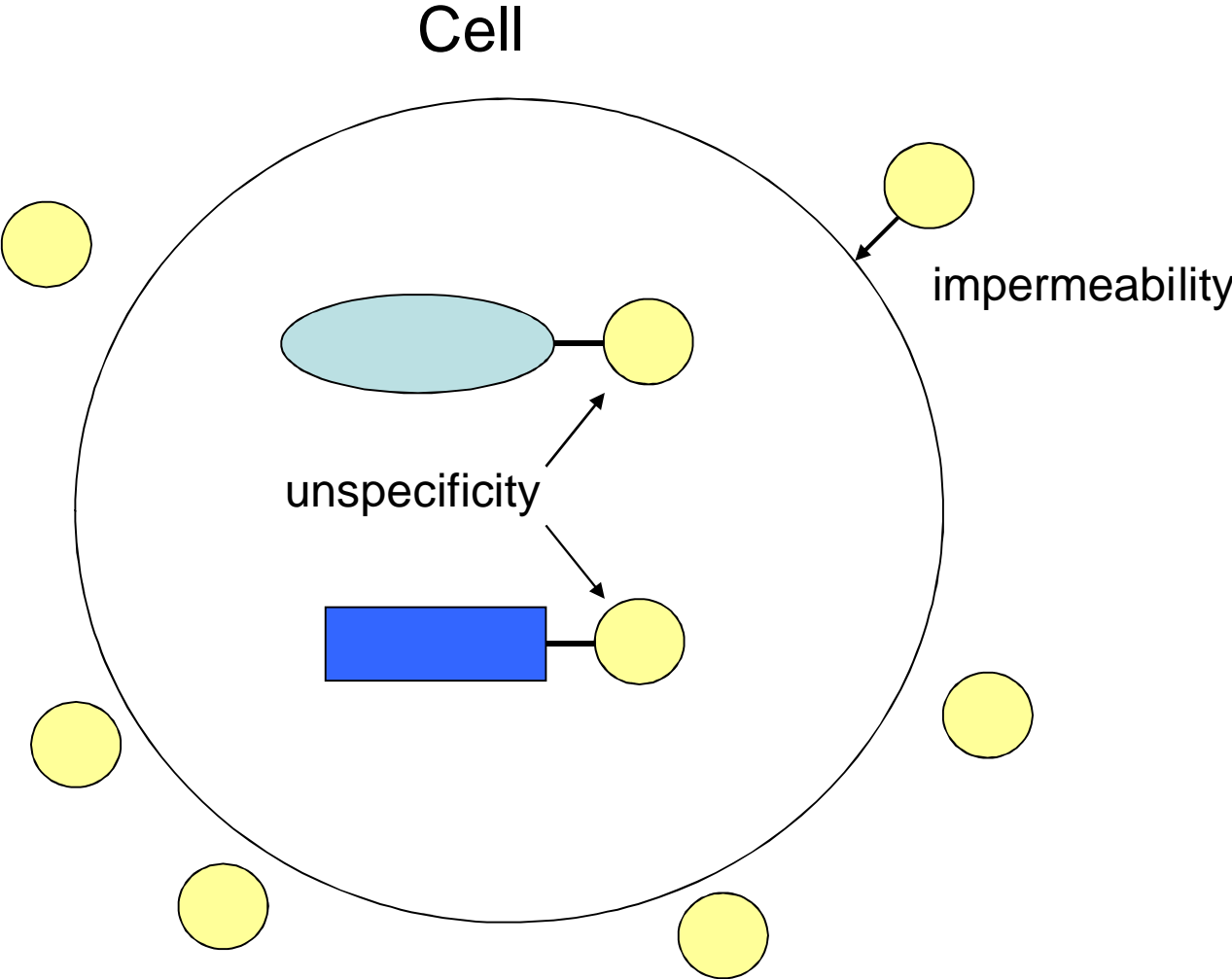
Compound microscope



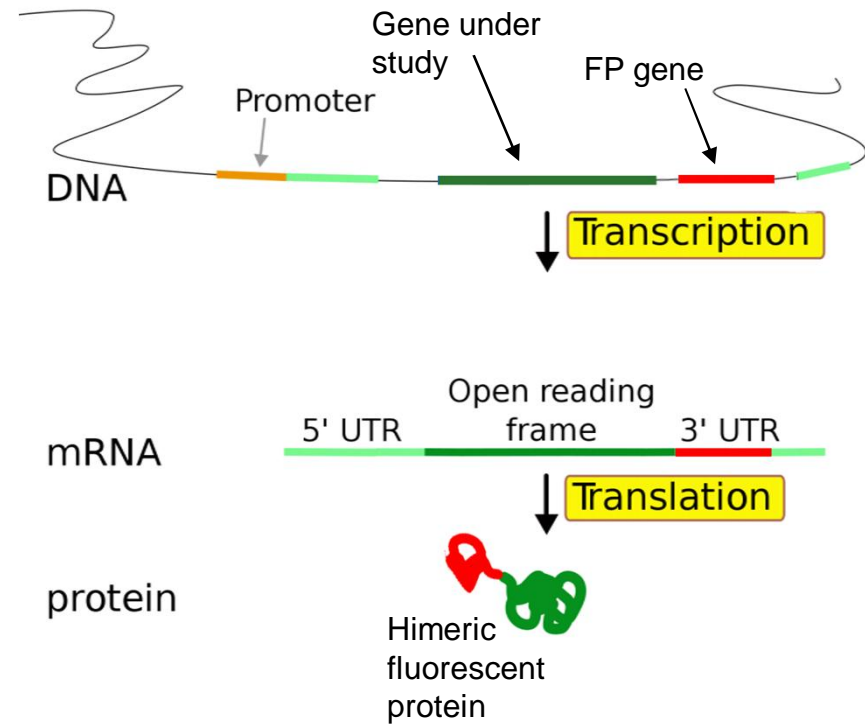
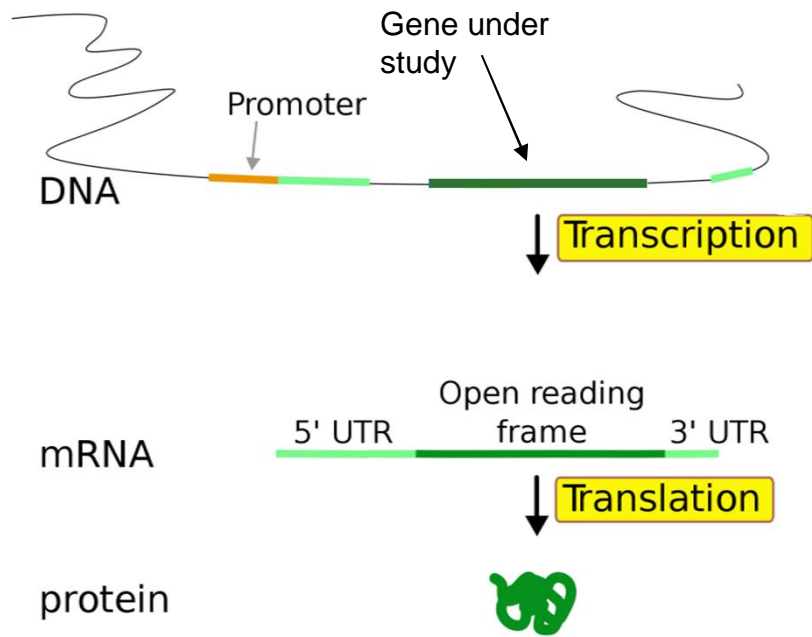
iMIC Digital Microscopes

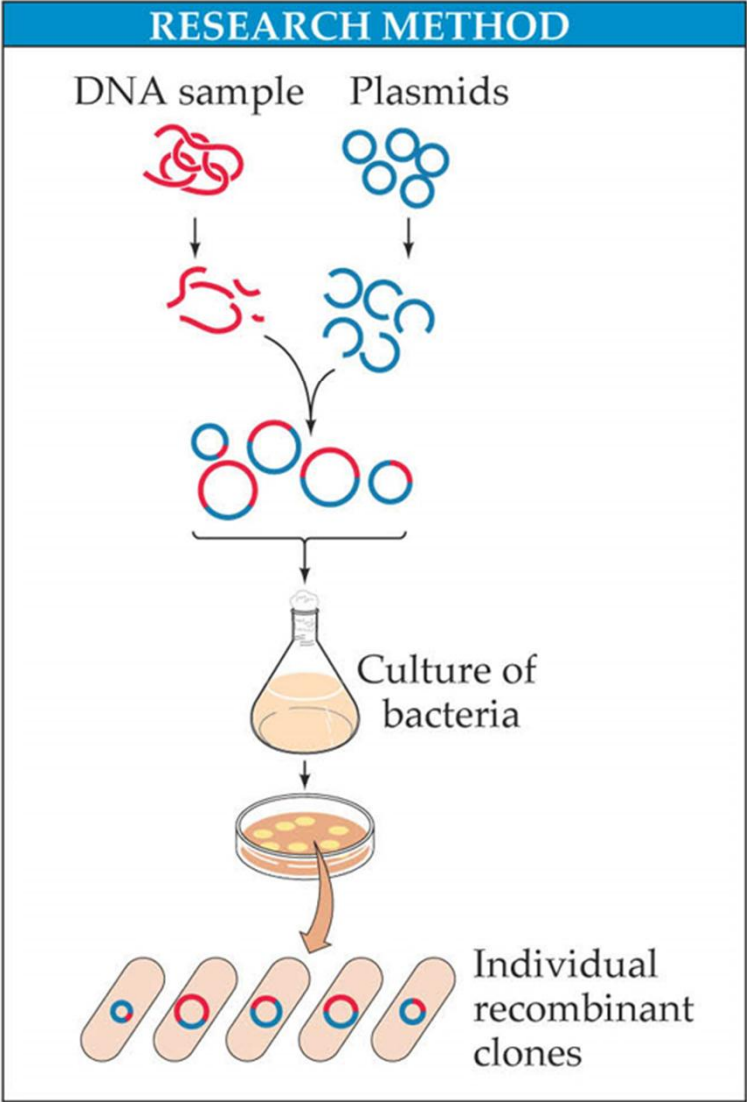


# Problems with fluorescent tags

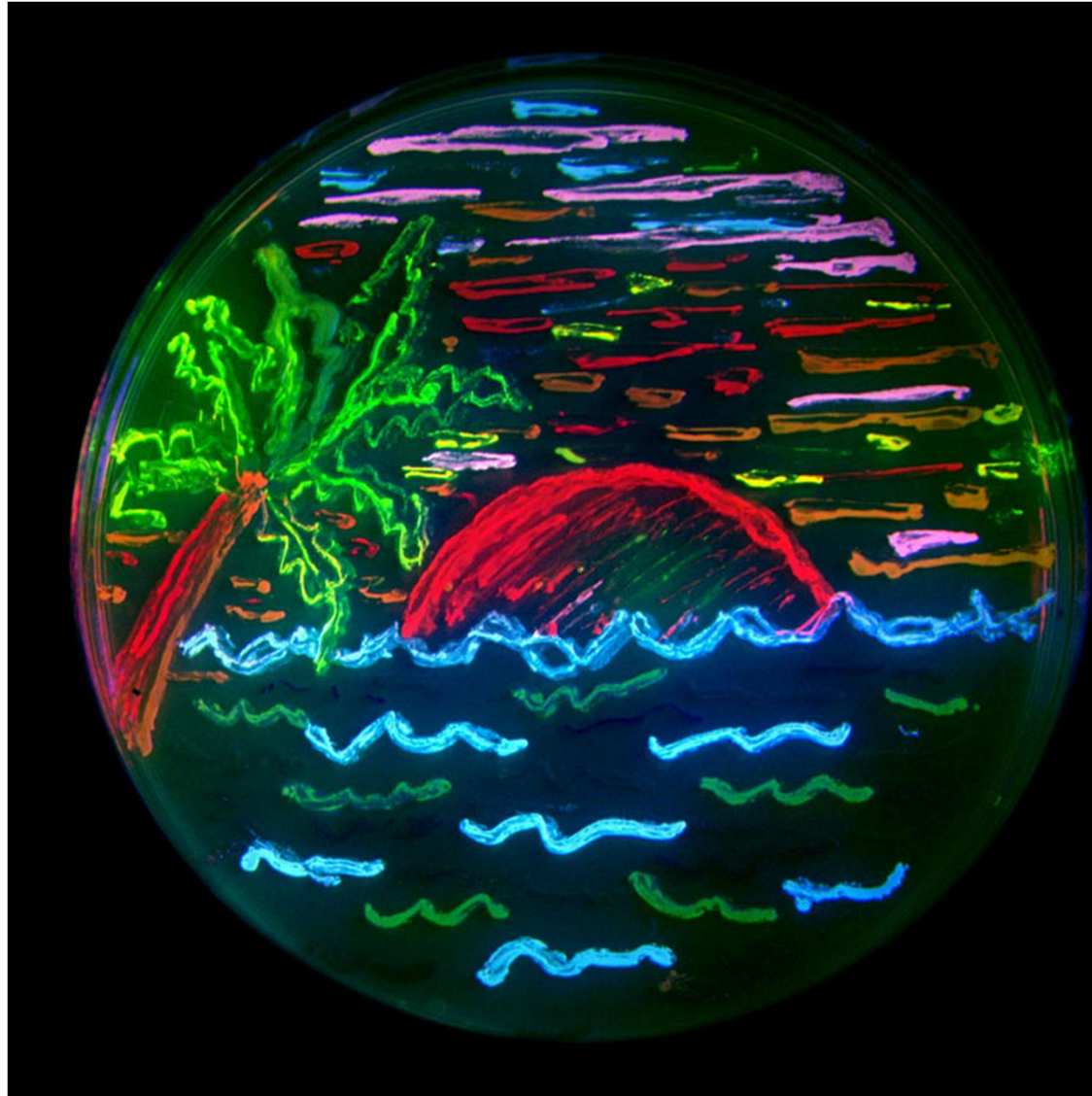


# Himeric gene expression

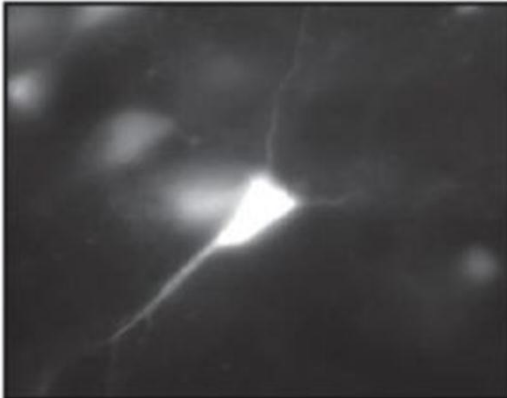
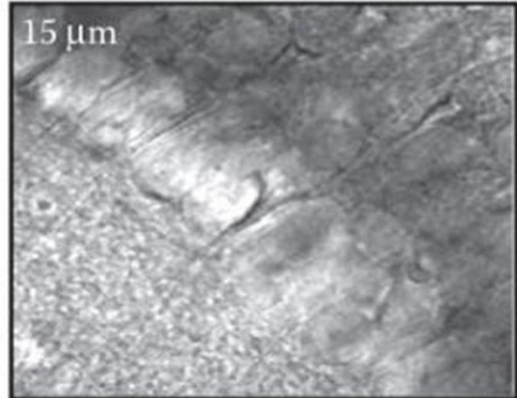
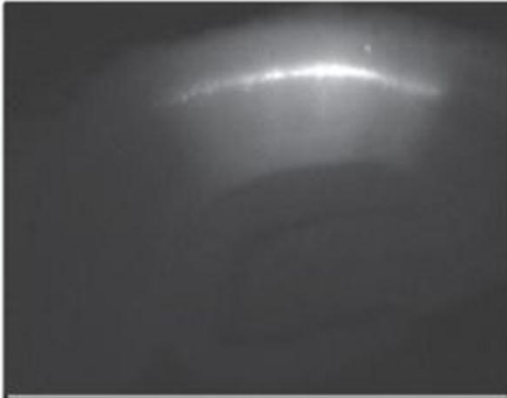
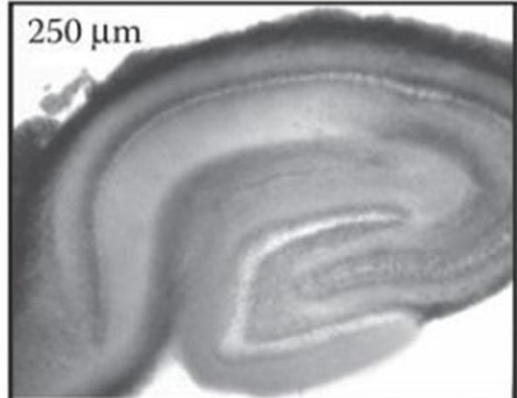




(GFP)



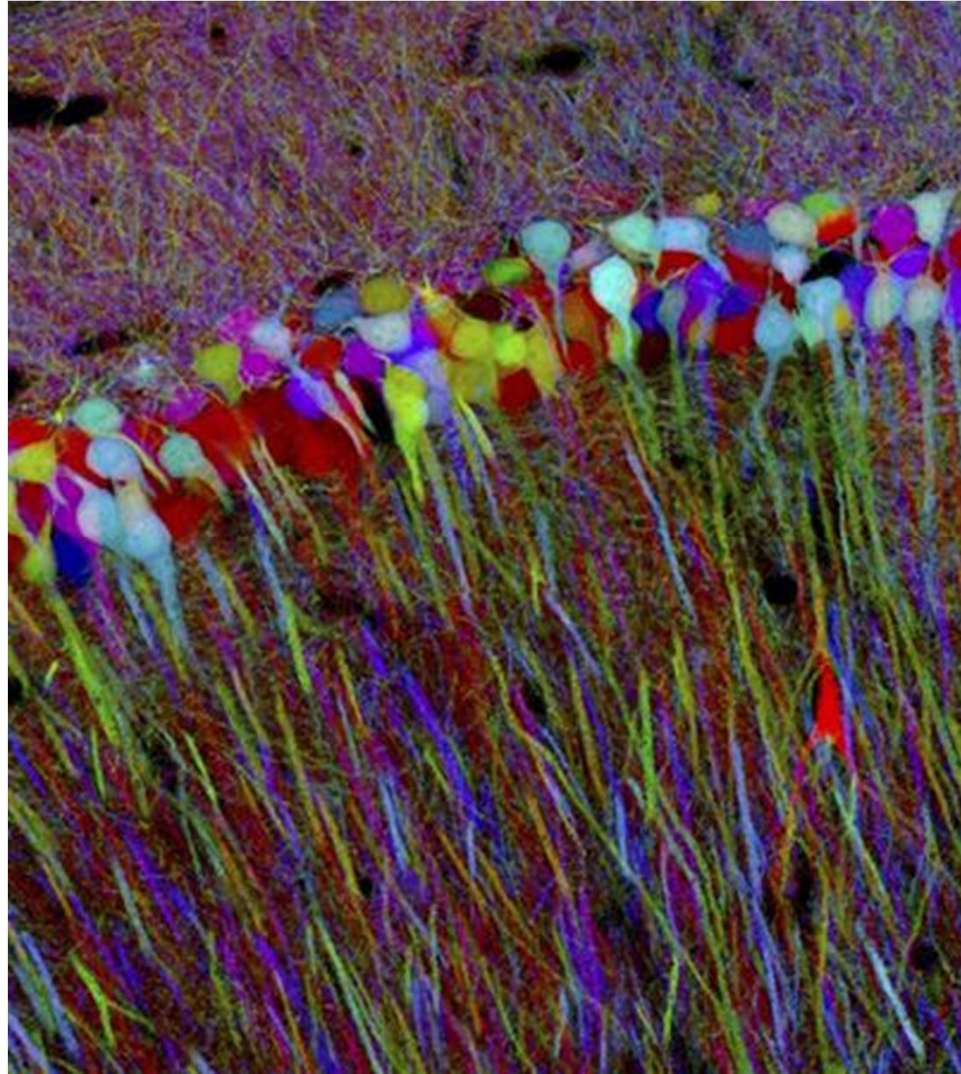
<http://www.conncoll.edu/ccacad/zimmer/GFP-ww/GFP-1.htm>



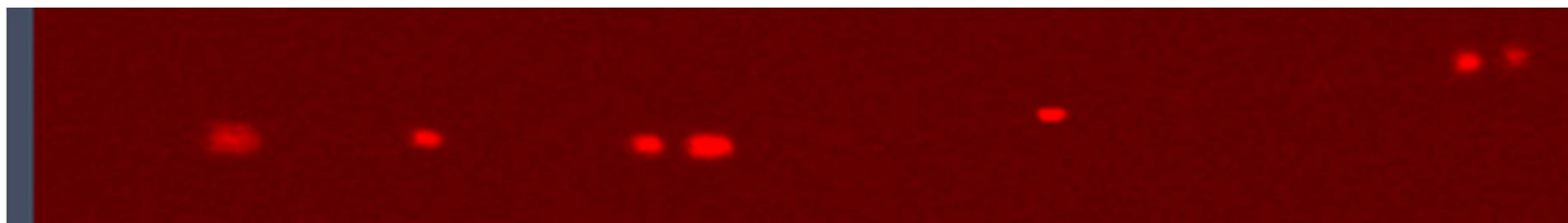


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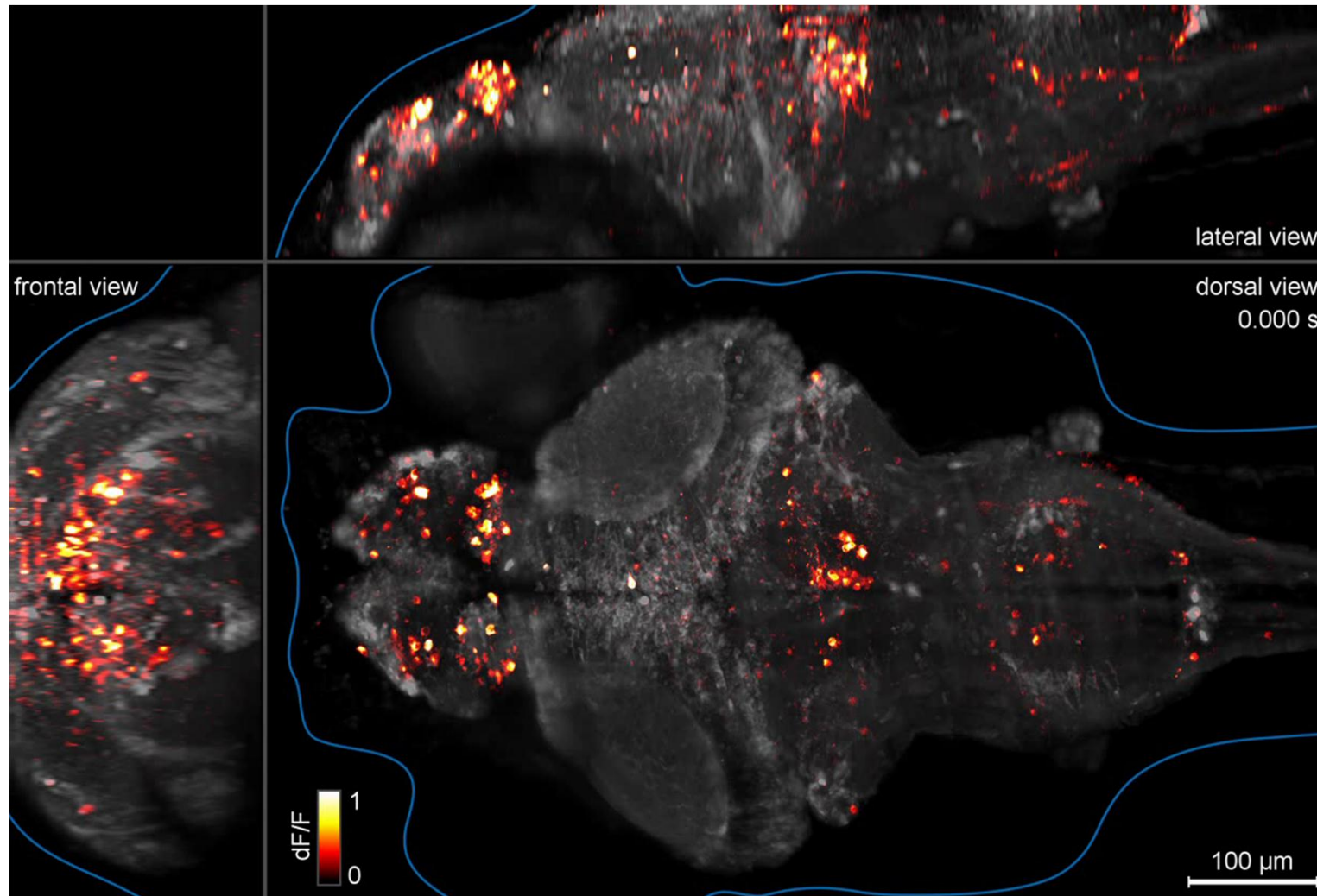
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# Mitochondrial dynamics in axons of hippocampal pyramidal neurons



# Whole-brain imaging of neuronal activity



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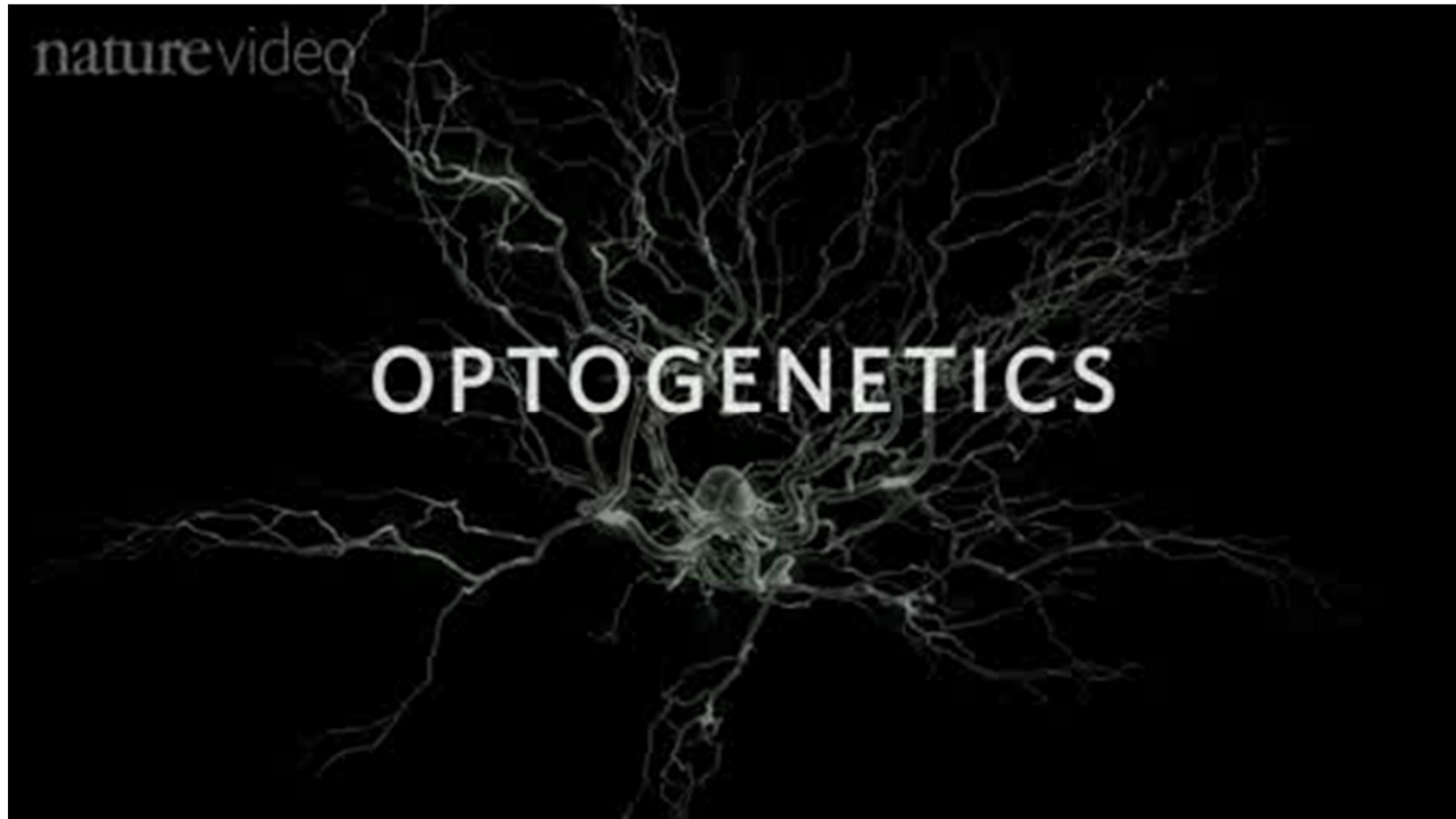
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## (Optogenetics)

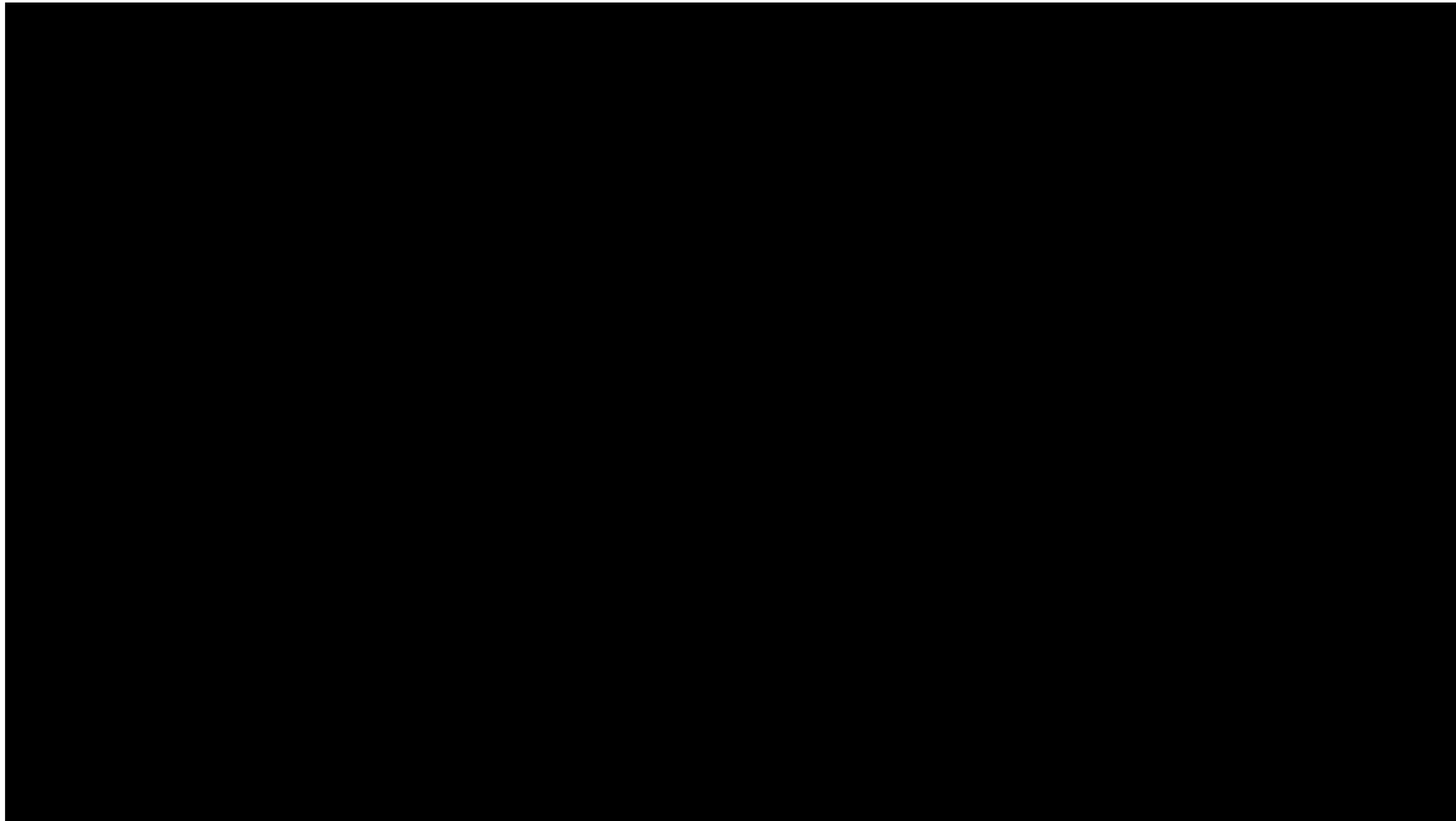
**Optogenetics** is the combination of genetic and optical methods to control specific events in targeted cells of living tissue, even within freely moving [mammals](#) and other animals, with the temporal precision ([millisecond](#)-timescale) needed to keep pace with functioning intact biological systems.

Photosensitive bacteria are founders of optogenetics



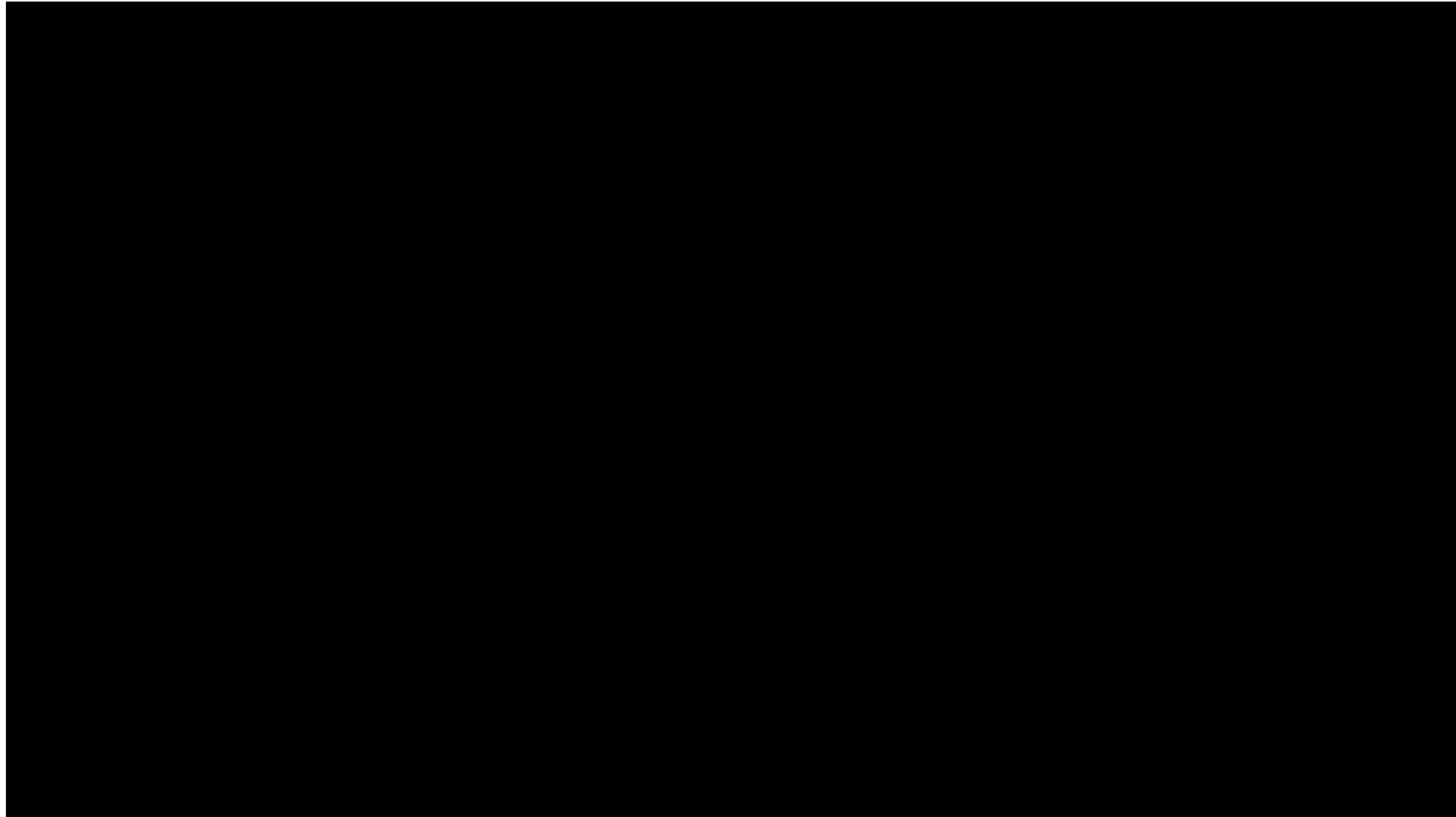
Nature web site

# Channelrhodopsin (ChR2) an electronic photoswitch exciting neurons

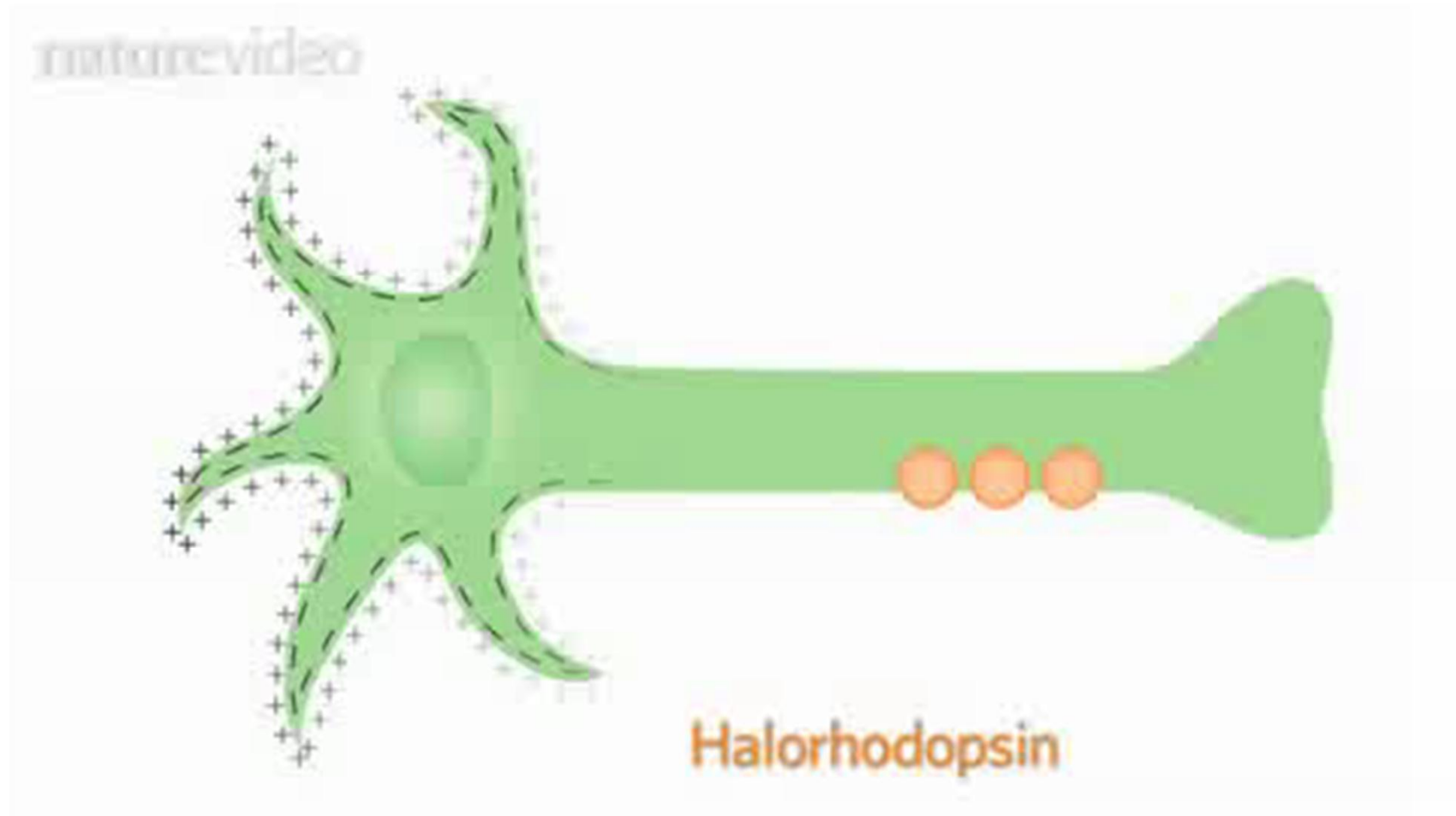




# Examples of how Channelrhodopsins can control cells and animal behavior



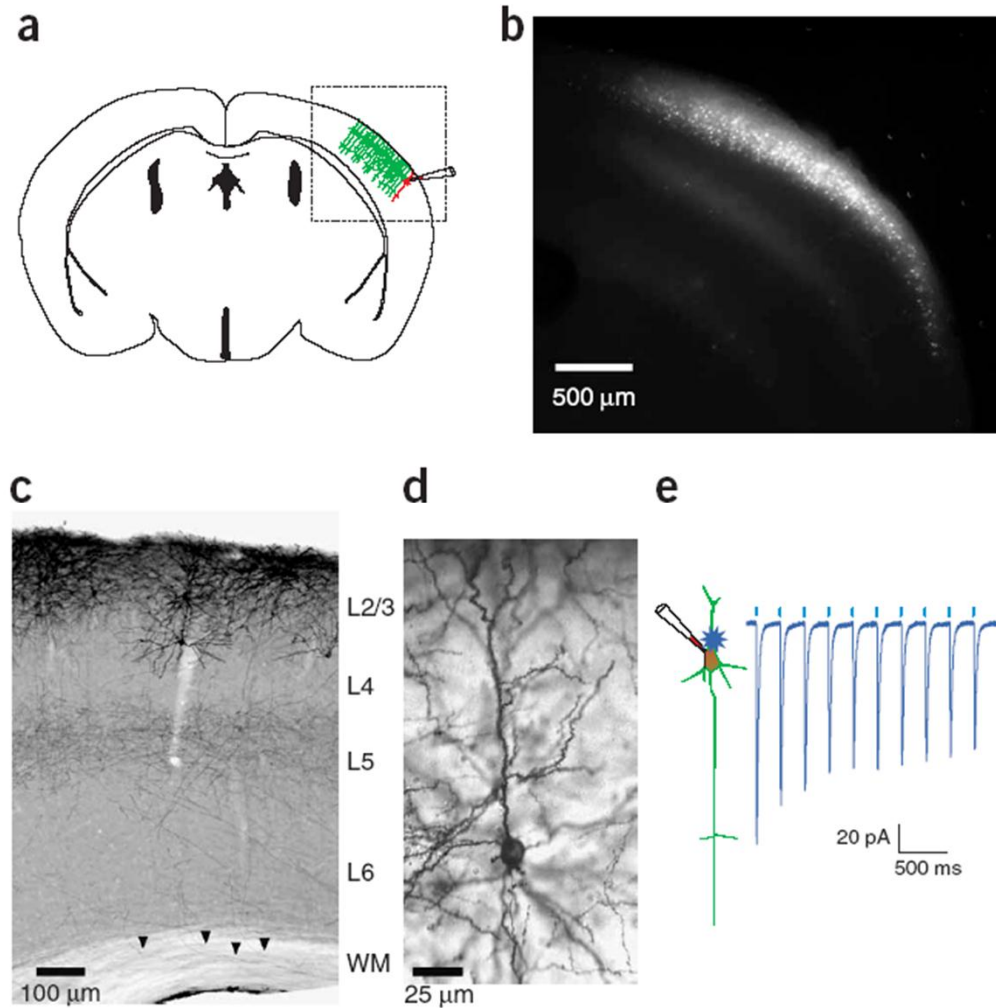
# Halorhodopsin (NpHR) - an electronic photoswitch inhibiting neurons



# Molecular-genetic approach to introduce optogenetic constructs

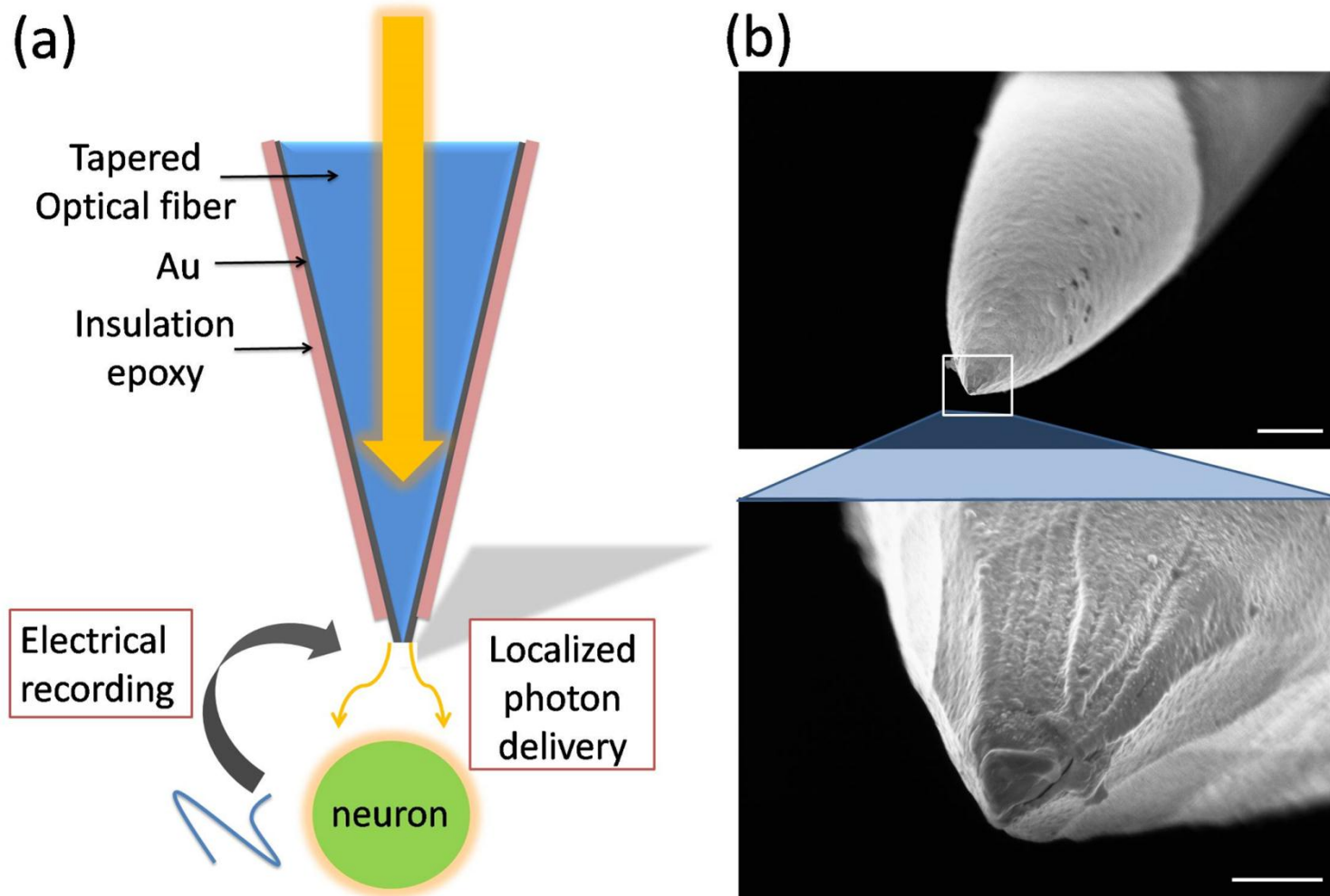


# Photostimulation of ChR2-positive neocortical L2/3 pyramidal neurons

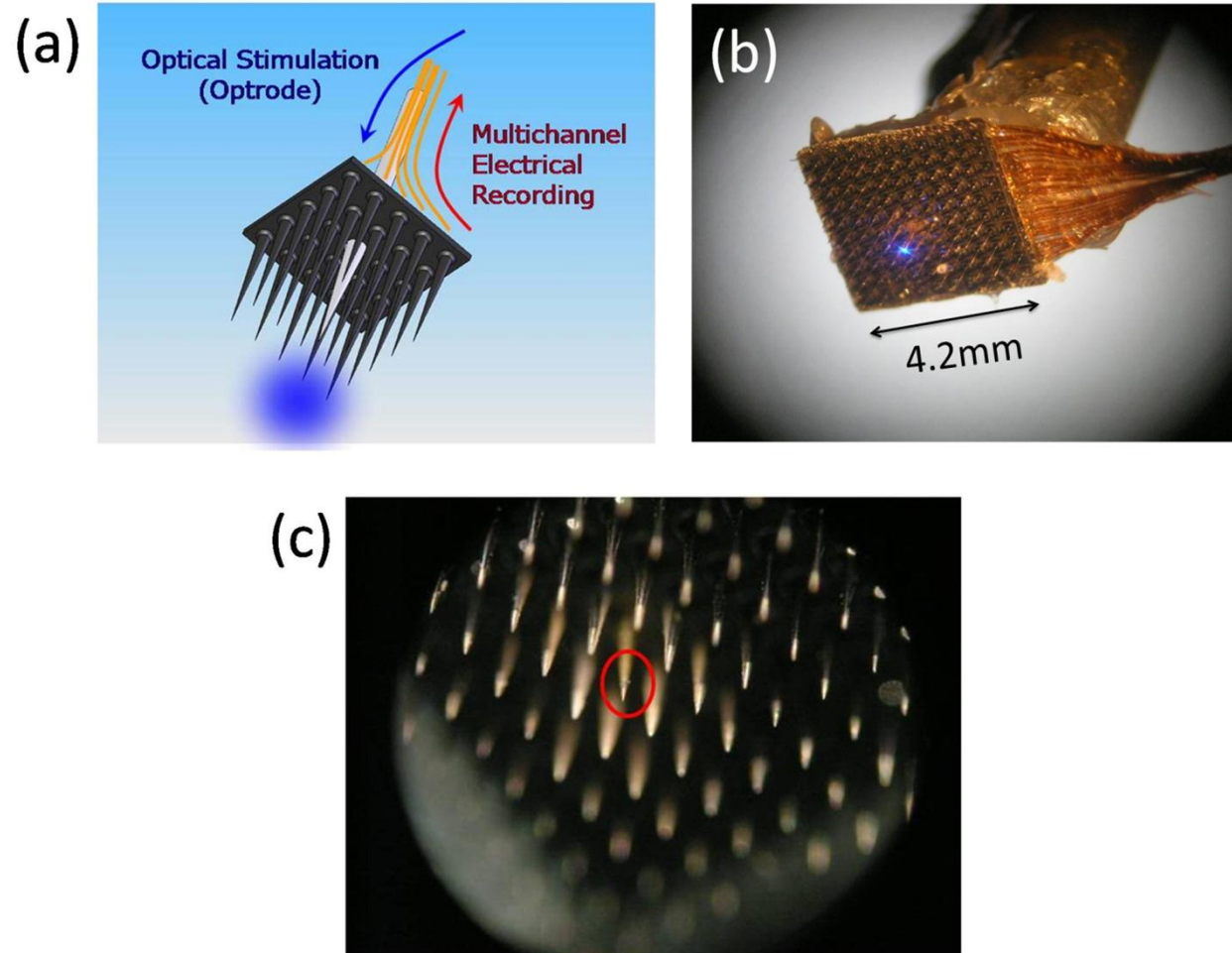


NATURE NEUROSCIENCE 10:663 2007.

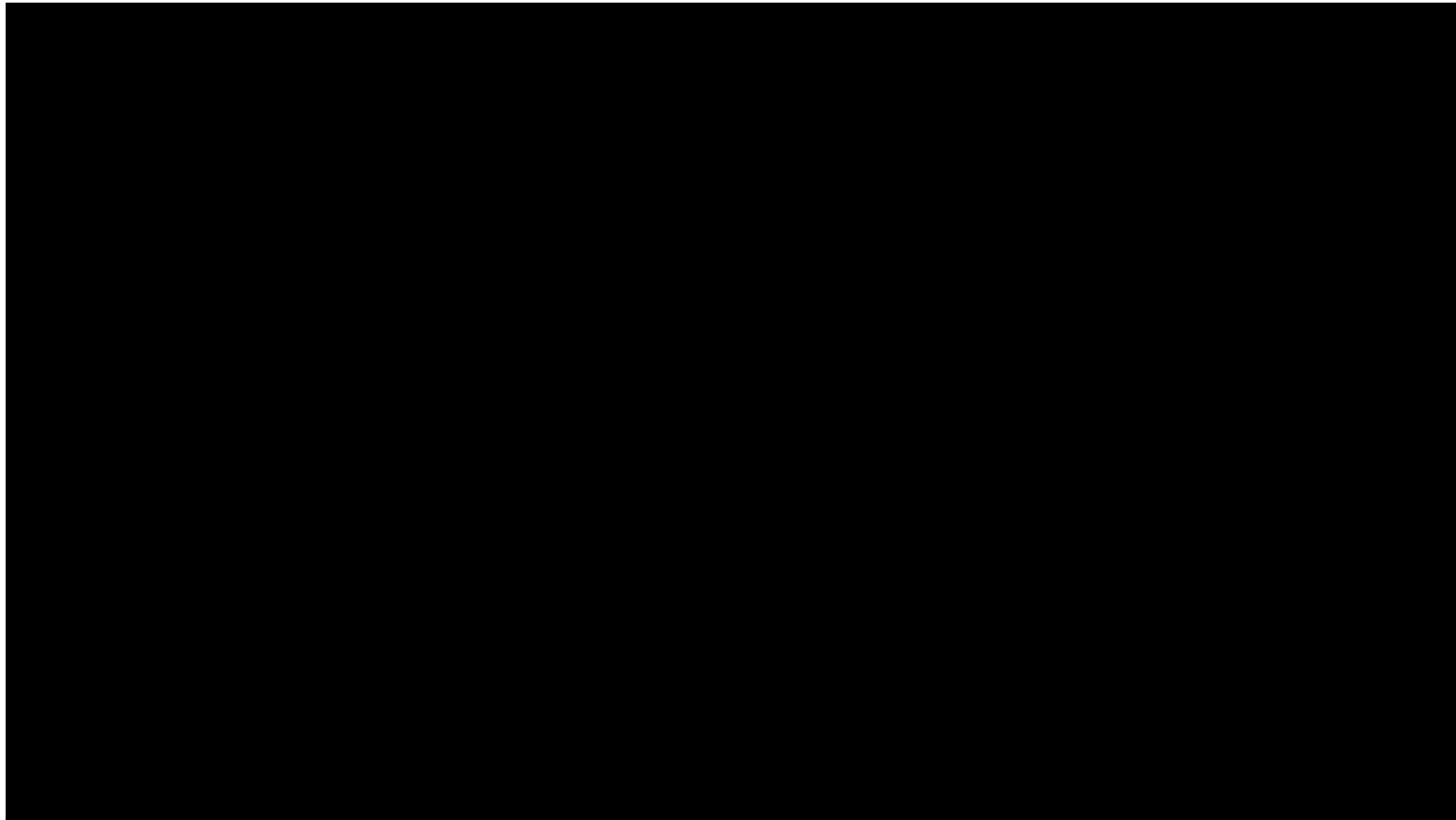
# Schematics and SEM images of optrode



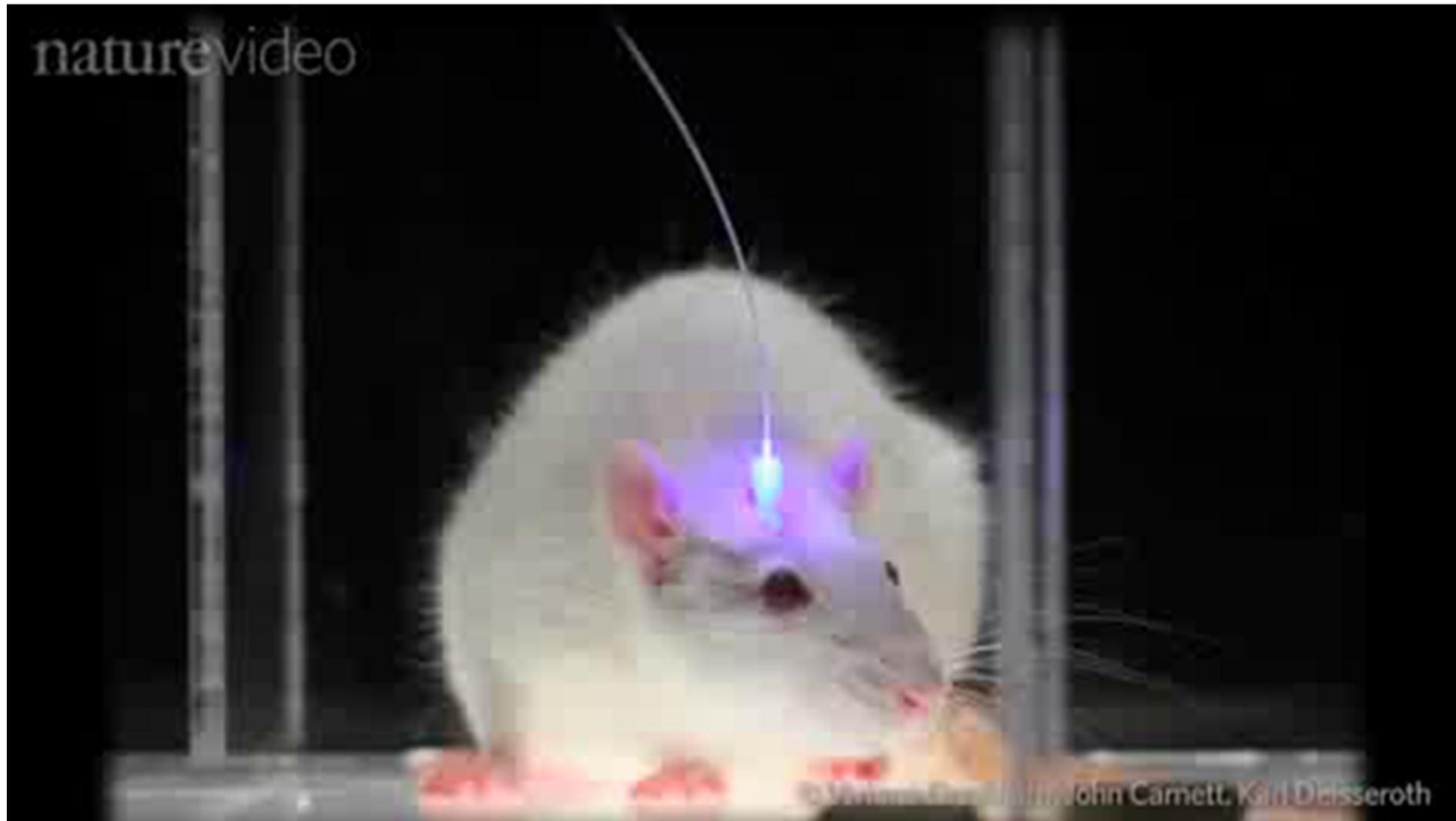
# Optrode array assembly



# ChR2 mouse

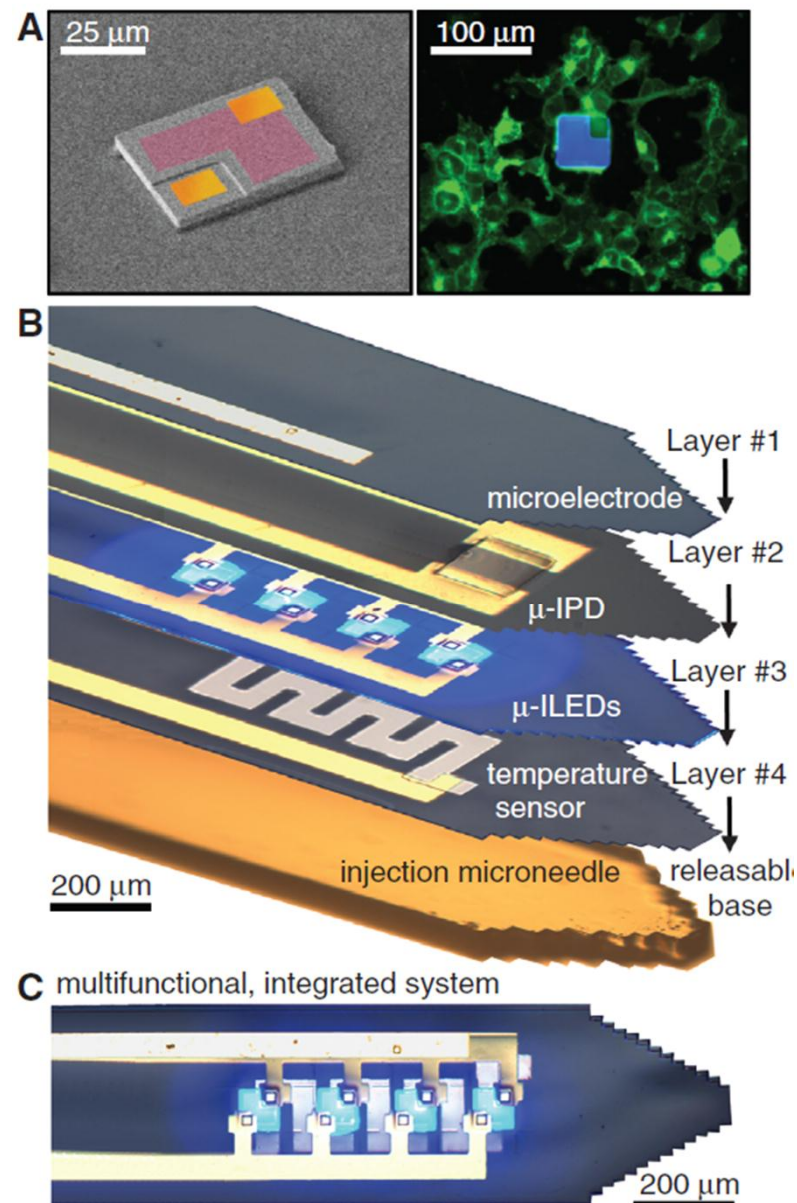


## Parkinson disease treatment by optogenetic approaches

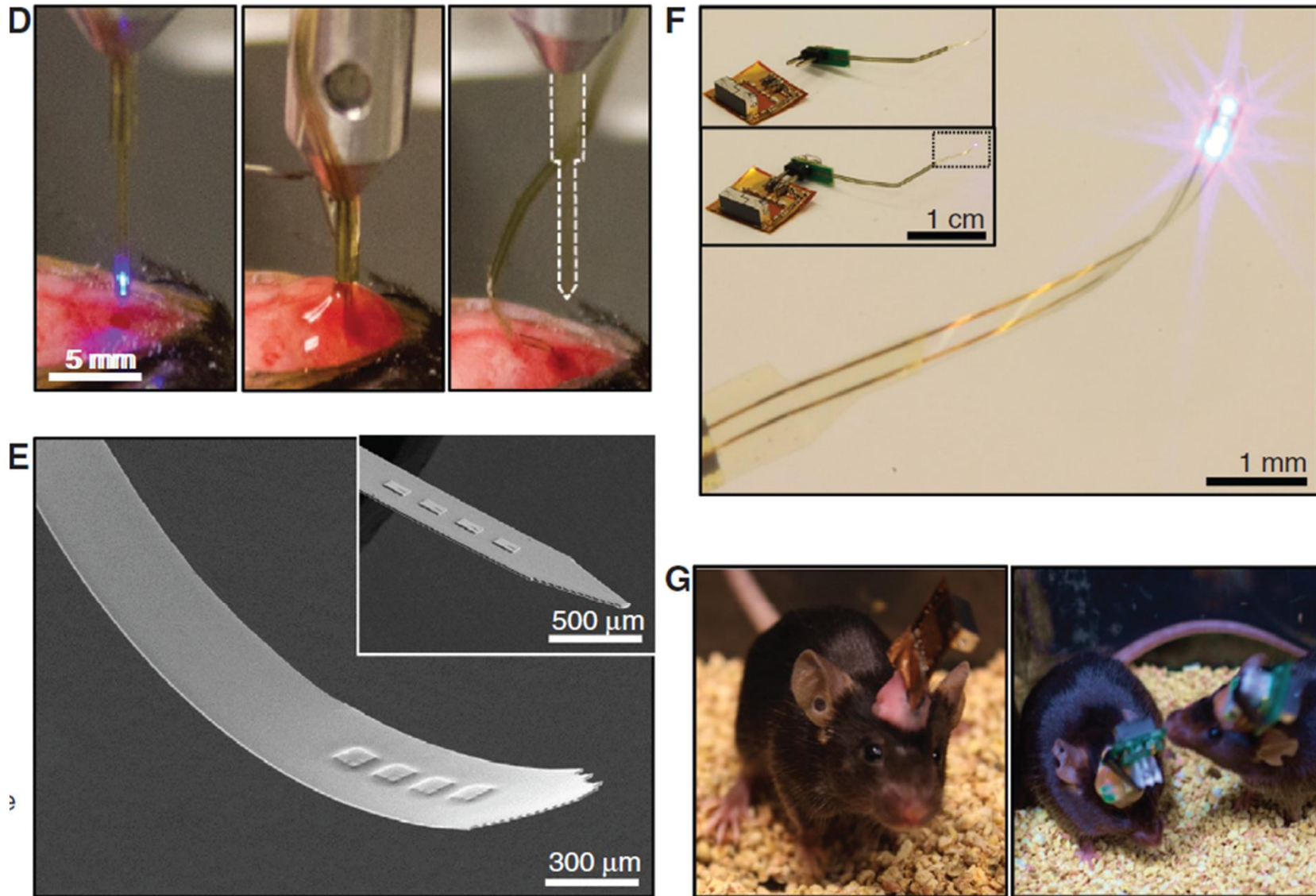




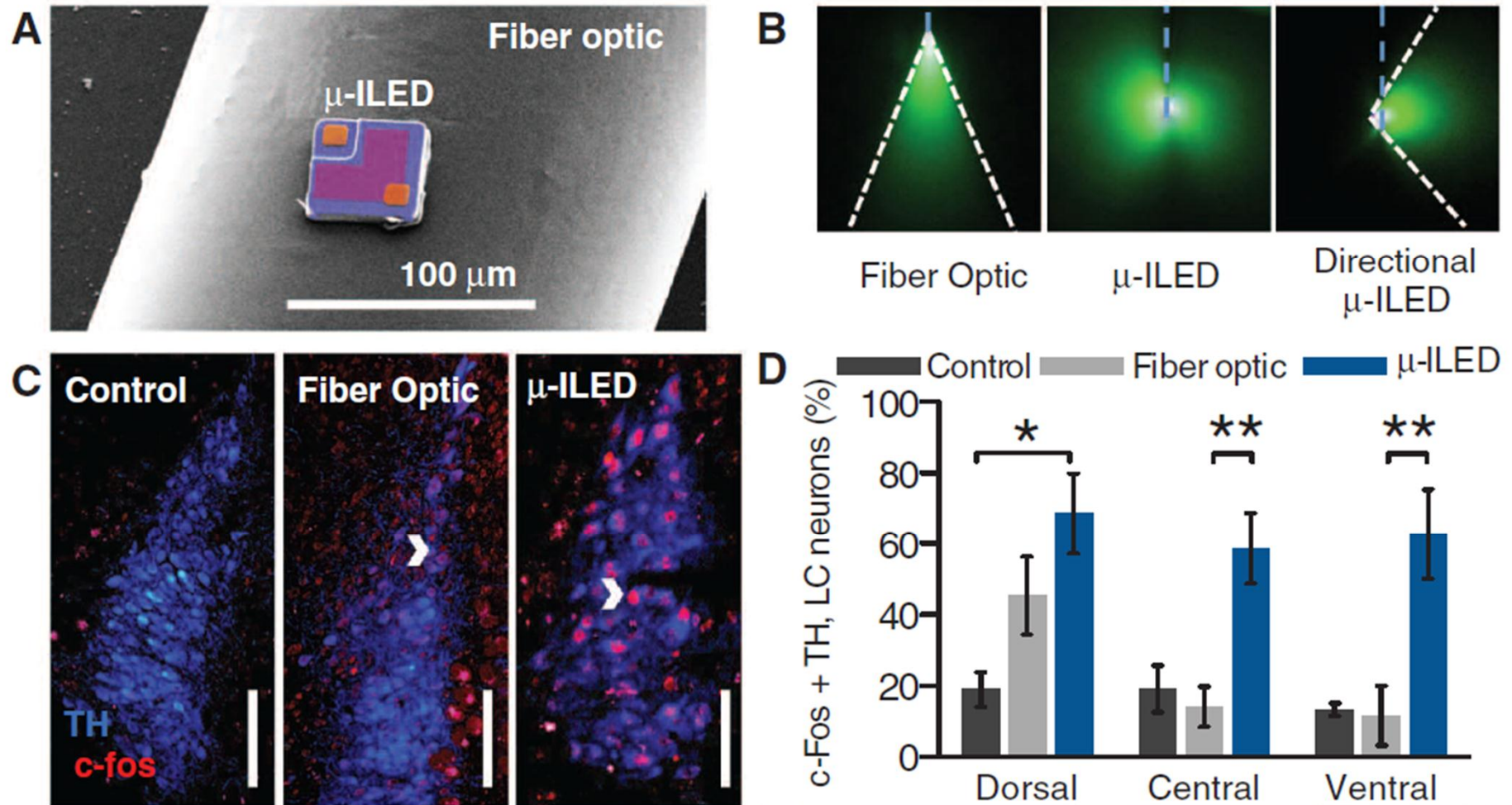
# Injectable, cellular-scale semiconductor devices



# Injectable, cellular-scale semiconductor devices



# $\mu$ -ILED devices improve spatial targeting



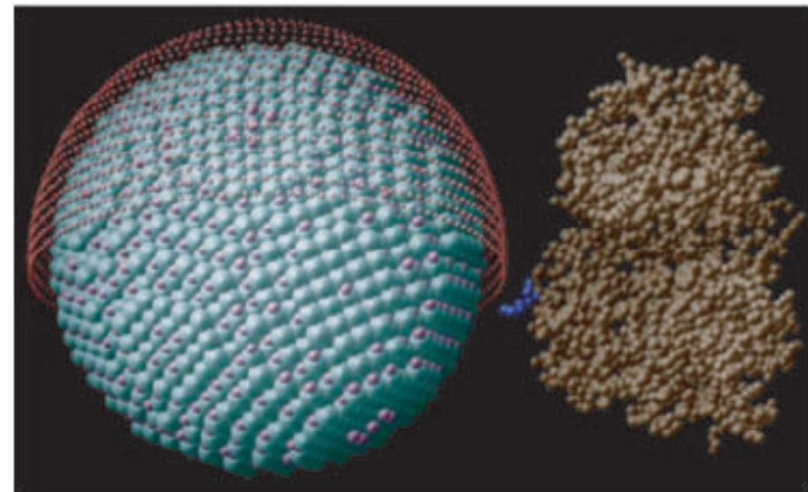
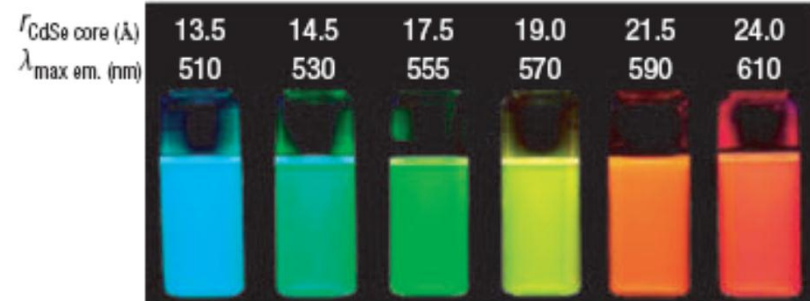
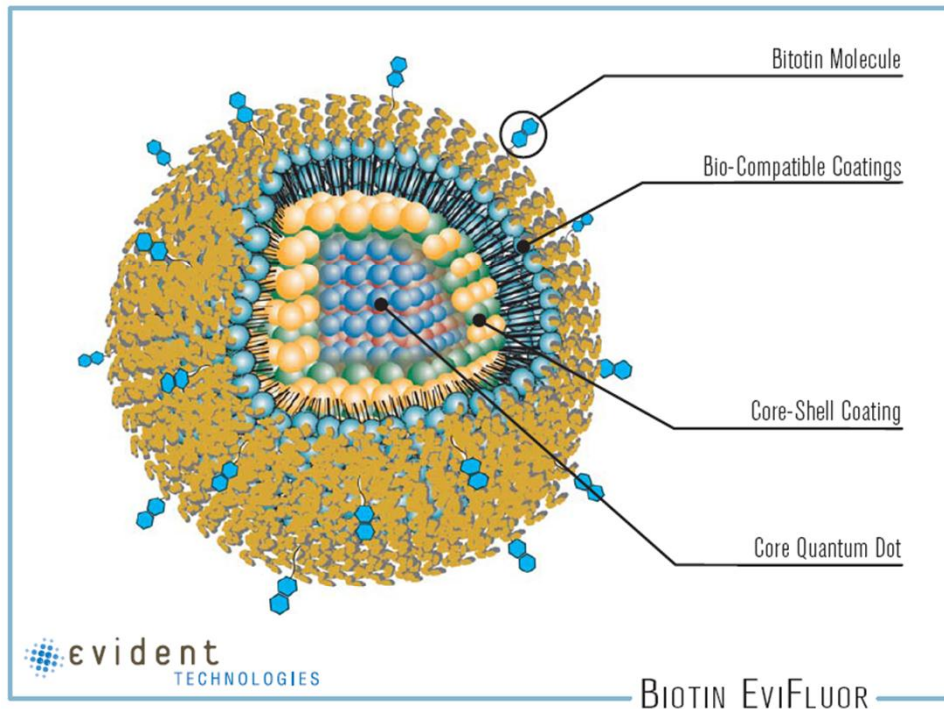
# Conclusions

- “ **Optogenetics** is a new experimental methodology based upon the combination of genetic and optical methods to control specific mainly electrical events in targeted cells of living tissue.
- “ **Optogenetics** allows to almost instantaneously switch on and off certain neuronal groups in order to study their functions and relationships with other cells of animal body.
- “ **Optogenetics** also allows to correct pathological changes of signaling processes being a prerequisite for treatment of many neurodegenerative disorders.

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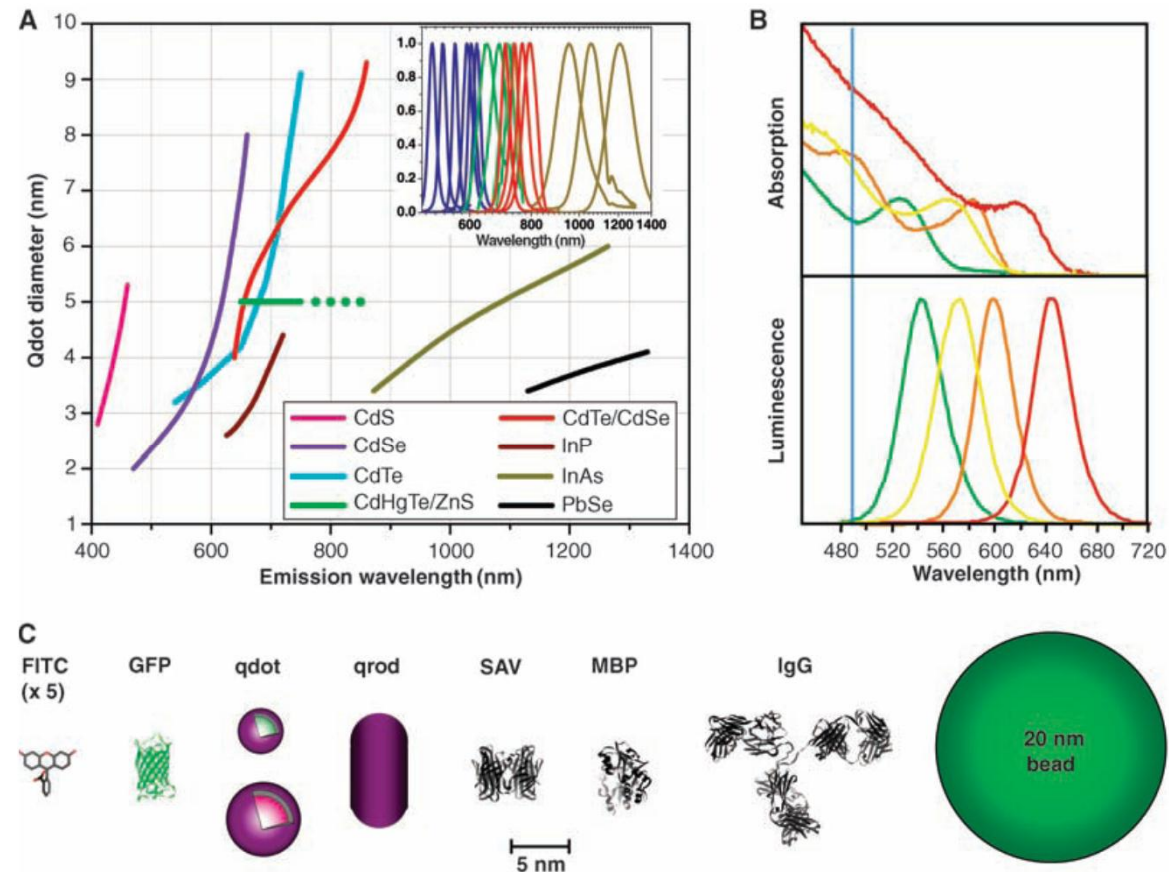




Nature Materials, 4:435, 2005

QDot as compared to MBP, a midsize protein ( $M_r \sim 44 \text{ kDa}$ ) with dimensions of  $30 \times 40 \times 65 \text{ \AA}$

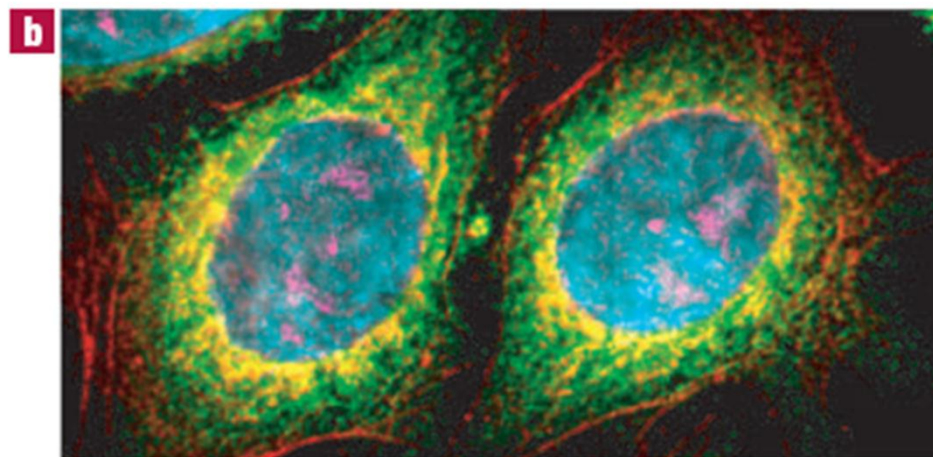
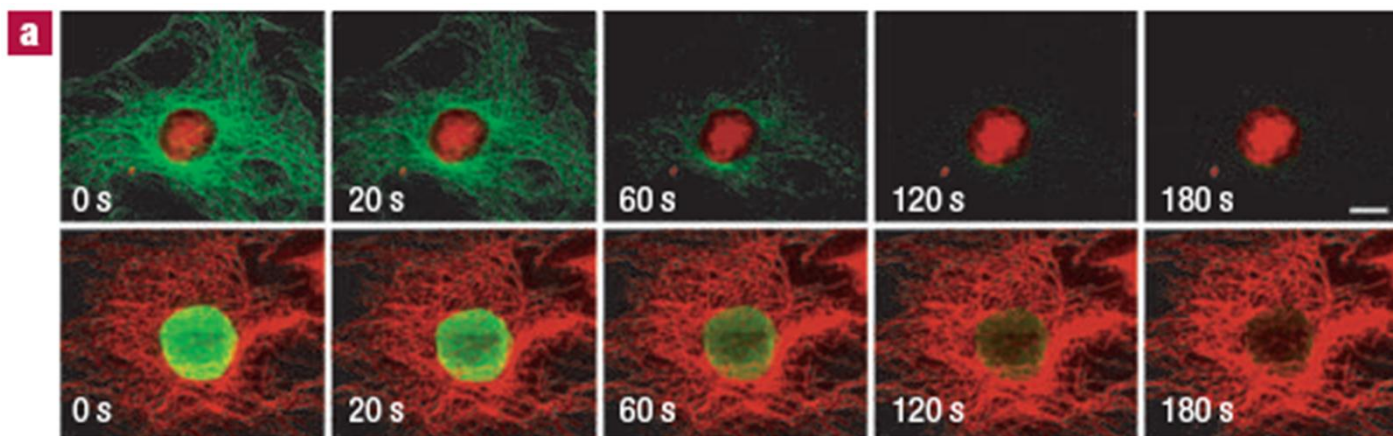
Evident technology web site



(A) Emission maxima and sizes of quantum dots of different composition. Quantum dots can be synthesized from various types of semiconductor materials (II-VI: CdS, CdSe, CdTe; III-V: InP, InAs; IV-VI: PbSe) characterized by different bulk band gap energies. The curves represent experimental data from the literature on the dependence of peak emission wavelength on qdot diameter. The range of emission wavelength is 400 to 1350 nm, with size varying from 2 to 9.5 nm (organic passivation/solubilization layer not included). All spectra are typically around 30 to 50 nm (full width at half maximum). Inset: Representative emission spectra for some materials. (B) Absorption (upper curves) and emission (lower curves) spectra of four CdSe/ZnS qdot samples. The blue vertical line indicates the 488-nm line of an argon-ion laser, which can be used to efficiently excite all four types of qdots simultaneously. (C) Size comparison of qdots and comparable objects. FITC, fluorescein isothiocyanate; GFP, green fluorescent protein; qdot, green (4 nm, top) and red (6.5 nm, bottom) CdSe/ZnS qdot; qrod, rod-shaped qdot; streptavidin (SAV), maltose binding protein (MBP); and immunoglobulin G (IgG).

Quantum Dot Corp. Web site.

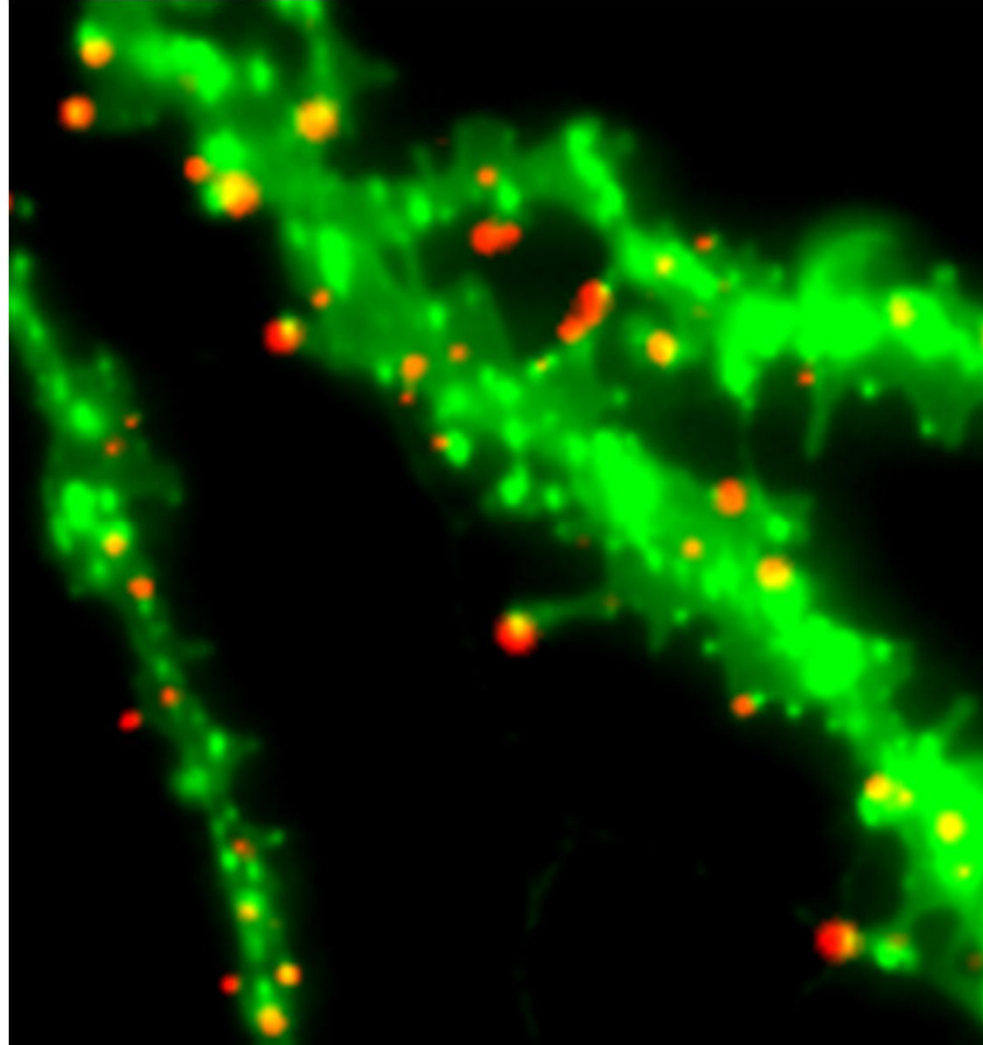
## QD resistance to photobleaching and multicolour labelling



**a**, Top row: Nuclear antigens were labelled with QD 630. streptavidin (red), and microtubules were labelled with AlexaFluor 488 (green) simultaneously in a 3T3 cell. Bottom row: Microtubules were labelled with QD 630. streptavidin (red), and nuclear antigens were stained green with Alexa 488. Continuous exposure times in seconds are indicated (Reprinted by permission of the Nature Publishing Group)<sup>34</sup>. Note the QD resistance to photobleaching under continuous illumination. **b**, Pseudocoloured image depicting five-colour QD staining of fixed human epithelial cells. Cyan corresponds to 655-nm Qdots labelling the nucleus, magenta 605-Qdots labelling Ki-67 protein, orange 525-Qdots labelling mitochondria, green 565-Qdots labelling microtubules and red 705-Qdots labelling actin filaments. Nature Materials, 4:435, 2005.

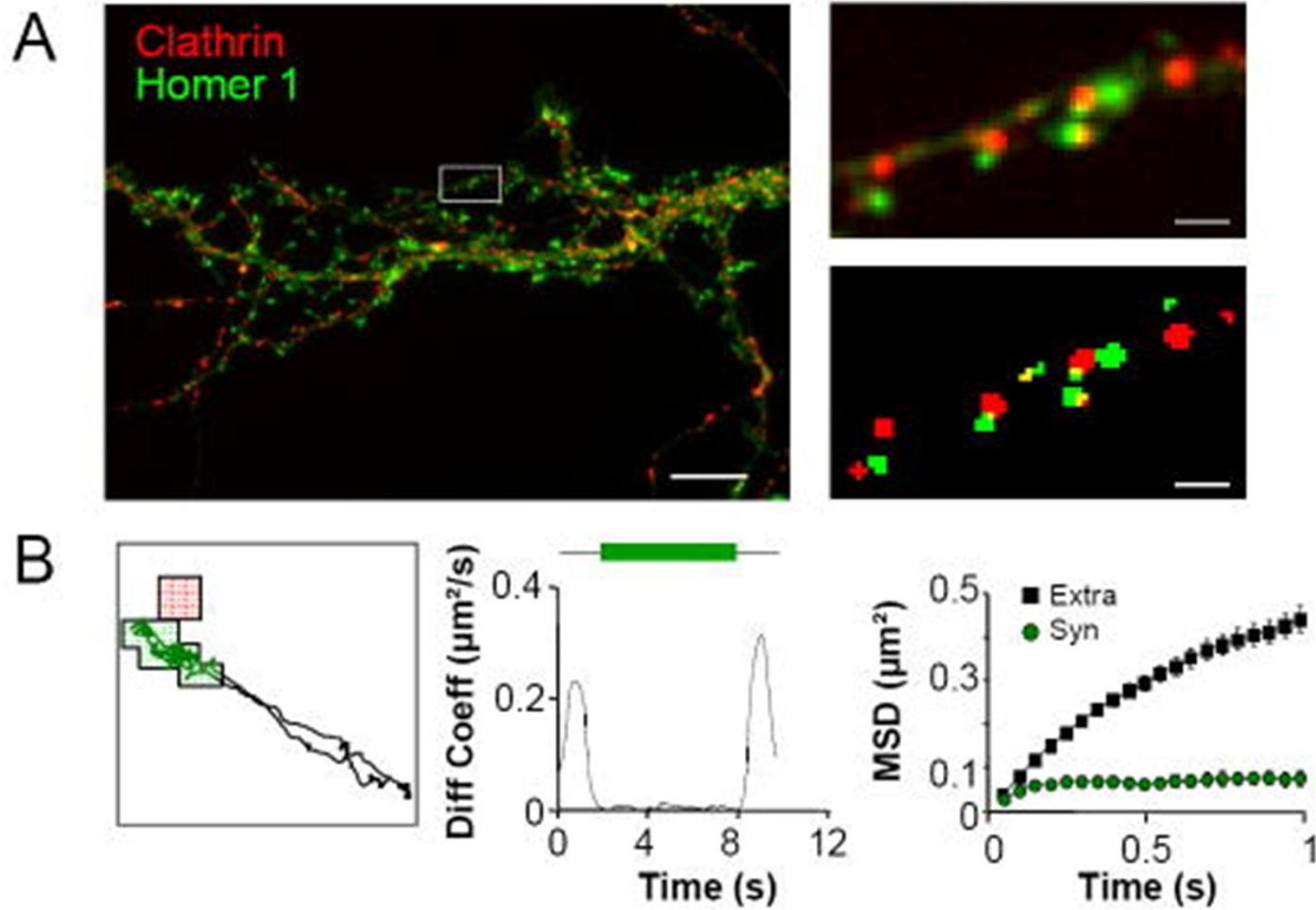


# AMPA



Neuron web site

# AMPA receptors are reversibly stabilized at synapses



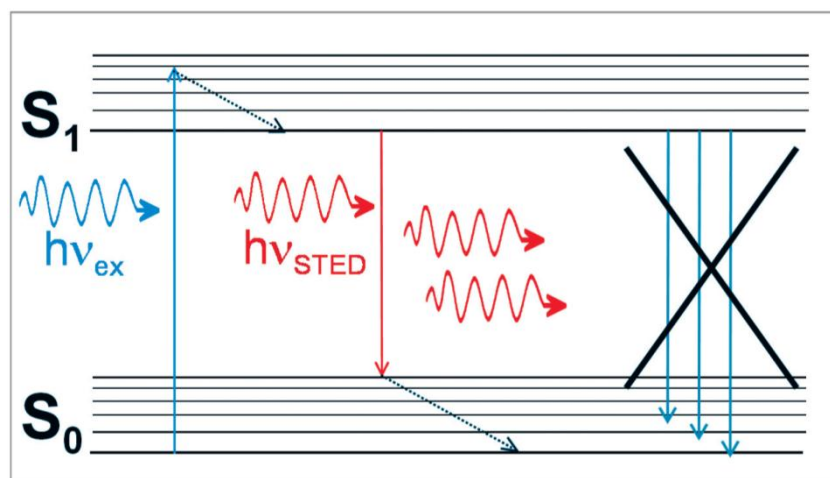
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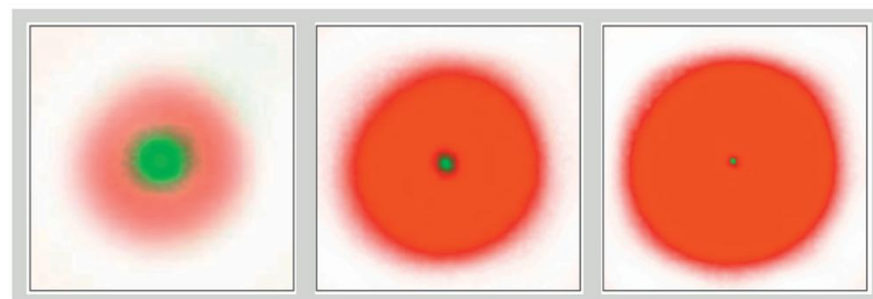
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# Stimulated Emission Depletion (STED) .



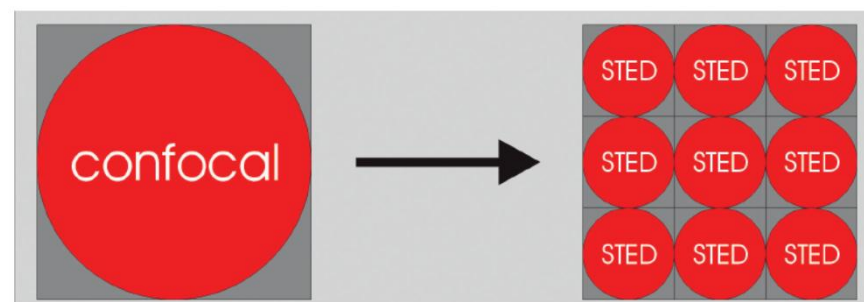
The effective fluorescent area (green) decreases with increasing depletion laser power (red)



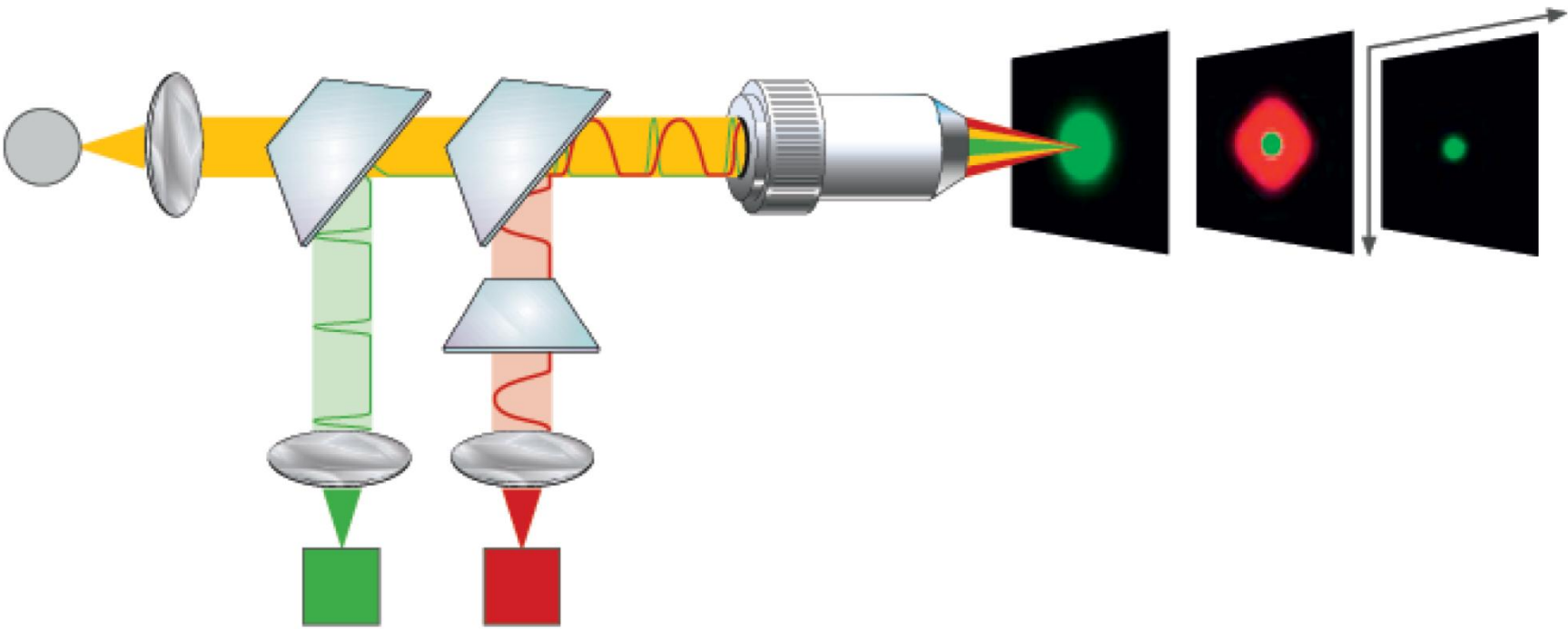
3-fold reduction of the scanning spot size in x and y yields 9-fold more accurate sampling

STED resolution:

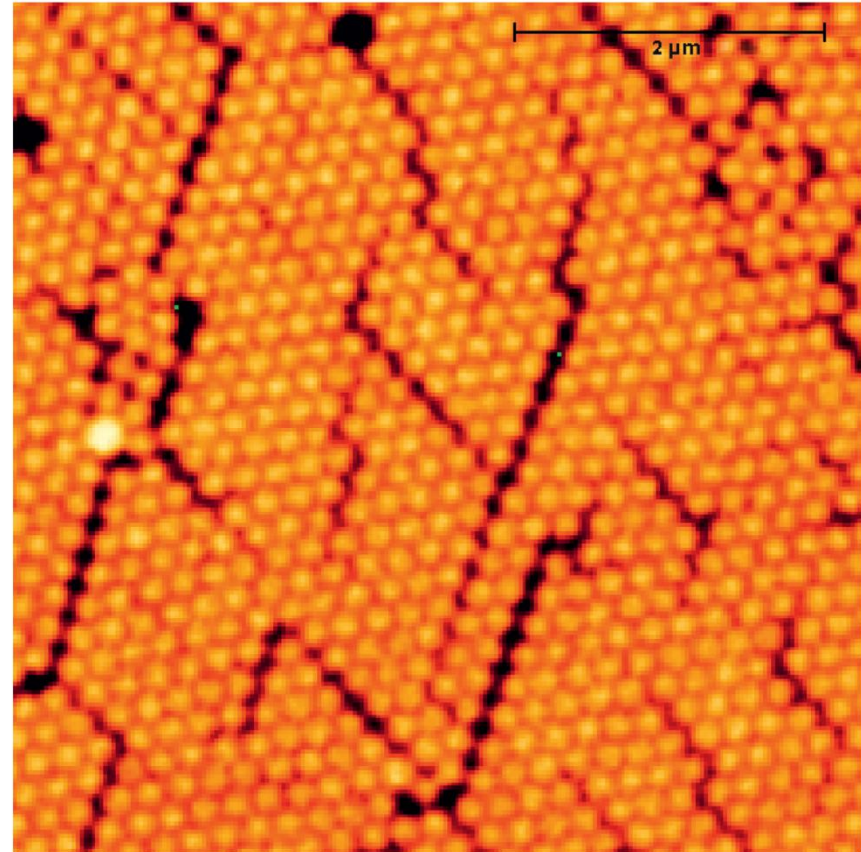
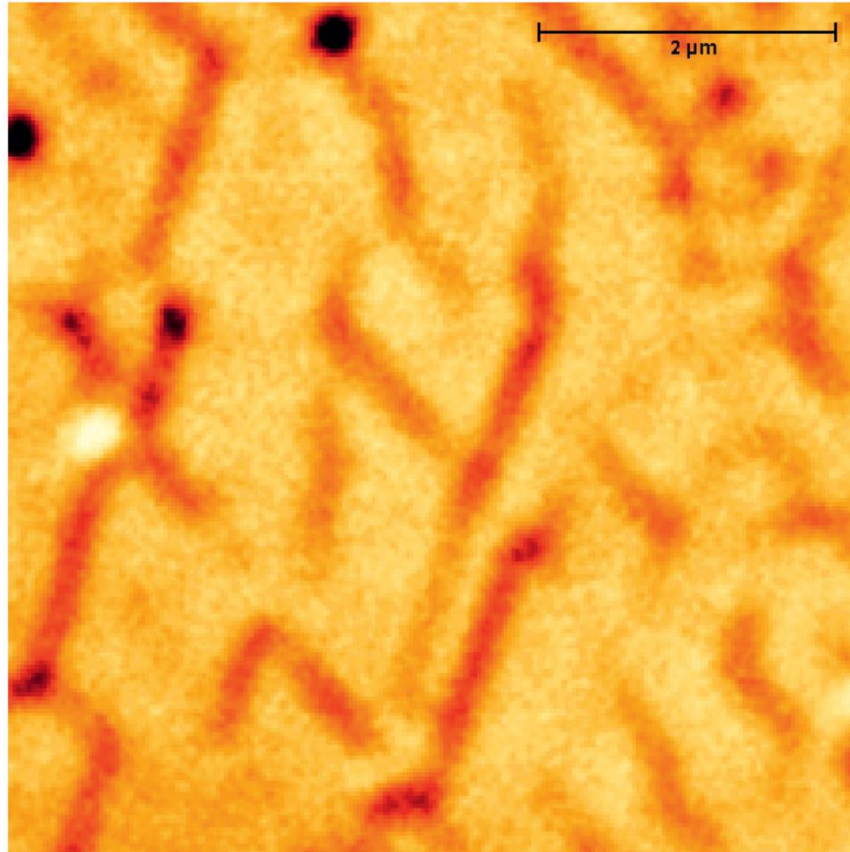
$$\Delta X \approx \frac{\lambda}{2n \sin \alpha \sqrt{1 + \frac{I}{I_s}}}$$



# Stimulated Emission Depletion (STED) .

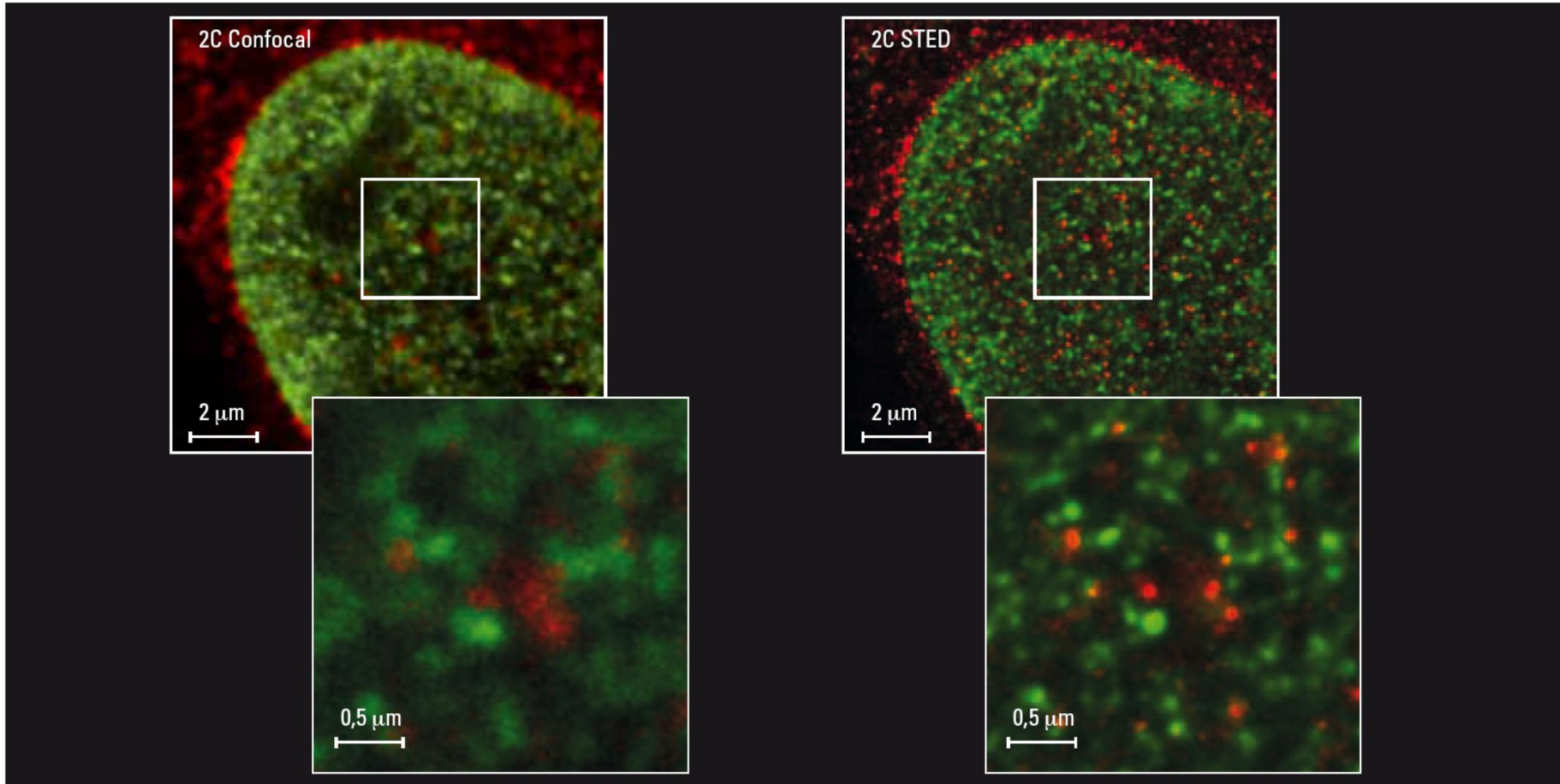


( ) STED ( )





( ) STED ( )

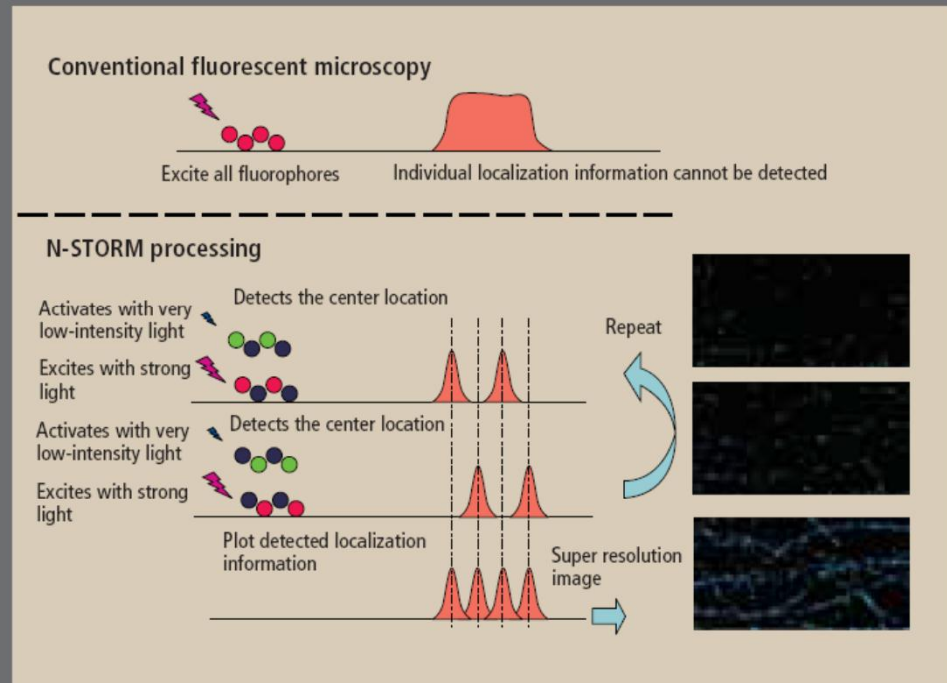


# STochastic Optical Reconstruction Microscopy (STORM)

## The Principle of N-STORM (STochastic Optical Reconstruction Microscopy)

STochastic Optical Reconstruction Microscopy (STORM) reconstructs a super resolution image by combining the high-accuracy localization information of each fluorophore in 3 spatial dimensions and multiple colors

N-STORM uses stochastic activation of relatively small numbers of fluorophore molecules using very low-intensity light. This low-level stochastic "activation" of discrete molecules enables high precision Gaussian fitting of each laterally. Additionally, taking advantage of an induced astigmatism via the special 3D-STORM optics, N-STORM localizes each molecule axially. Computationally combining molecular coordinates in 3 dimensions results in high contrast 3D images of the nanoscopic world with molecular specificity.





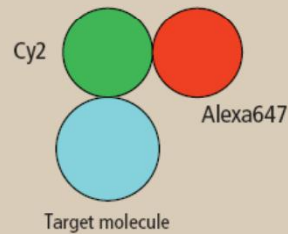
# STORM

## Dedicated fluorescent dyes

N-STORM uses dedicated fluorescent dye pairs containing an "activator" (relatively short wavelength excitation) and a "reporter" (relatively long wavelength excitation), which enables various color combinations, facilitating true multi-channel super resolution.

### Dyes for N-STORM

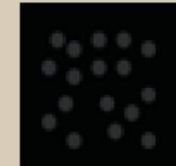
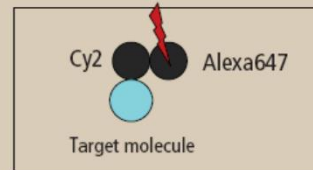
Dye for activation    Dye for image capturing



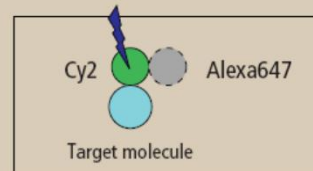
Dye for activation	Dye for image capturing
Alexa405	Alexa647
Cy2	Alexa647
Cy3	Alexa647

A dye for N-STORM consists of a shorter-wavelength dye for activation and a longer-wavelength dye for image capturing. Creation of two color super resolution images is possible with pairs of dye.

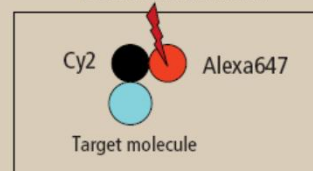
STEP 1 Inactivates all molecules



STEP 2 Activates Alexa647 by irradiating Cy2 with low-intensity light



STEP 3 Excite Alexa647 with strong light and capture images of localization information



Repeat more than 1,000 times

# N-SIM and N-STORM

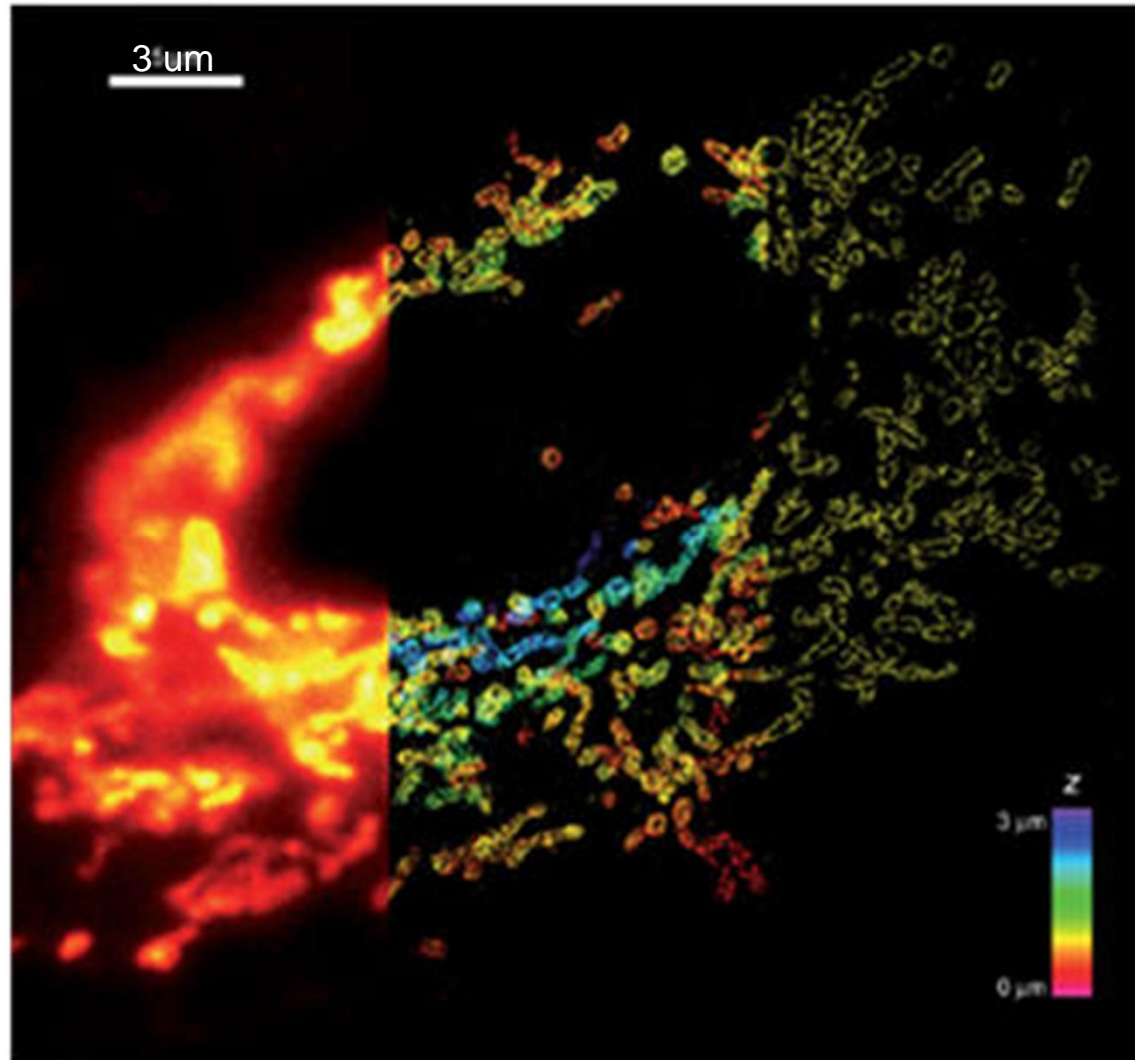
## Specifications

	N-SIM	N-STORM
XY resolution	Approx. 100nm (up to 85nm: theoretical, in TIRF-SIM mode, 488nm excitation)	Approx. 20nm
Z-axis resolution	Approx. 300nm	Approx. 50nm
Image acquisition time	Up to 0.6 sec./frame (TIRF-SIM/2D-SIM) Up to 1 sec. (3D-SIM) (needs more 1-2 sec. for calculation)	10 minutes or more
Imaging mode	TIRF-SIM (TIRF XY super resolution) 2D-SIM (XY super resolution, up to 3 $\mu$ m deep) 3D-SIM (XYZ super resolution, up to 20 $\mu$ m deep)	TIRF-STORM 3D STORM
Multi-color imaging	Up to 5 colors	2 colors simultaneously
Compatible Laser	Standard: 488nm, 561nm Option: 457nm, 515nm, 532nm	405nm, 457nm, 561nm, 647nm
Compatible microscopes	Motorized inverted microscope ECLIPSE Ti-E Perfect Focus System Motorized XY stage with encoders Piezo Z stage	
Objectives	CFI Apo TIRF 100 $\times$ oil (NA1.49) CFI Plan Apo IR 60 $\times$ WI (NA1.27)	CFI Apo TIRF 100x oil (NA 1.49) CFI Plan Apo VC 100x oil (NA 1.40)
Camera	Andor Technology iXon DU897 EMCCD camera	
Software	NIS-Elements Ar/ NIS-Elements C (with confocal microscope A1)	NIS-Elements Ar/ NIS-Elements C (with confocal microscope A1) Both need the NIS-A STORM Analysis
Operation conditions	25 $^{\circ}$ C $\pm$ 0.5 $^{\circ}$ C	20 $^{\circ}$ C to 25 $^{\circ}$ C ( $\pm$ 0.5 $^{\circ}$ C )

# Nikon STORM System

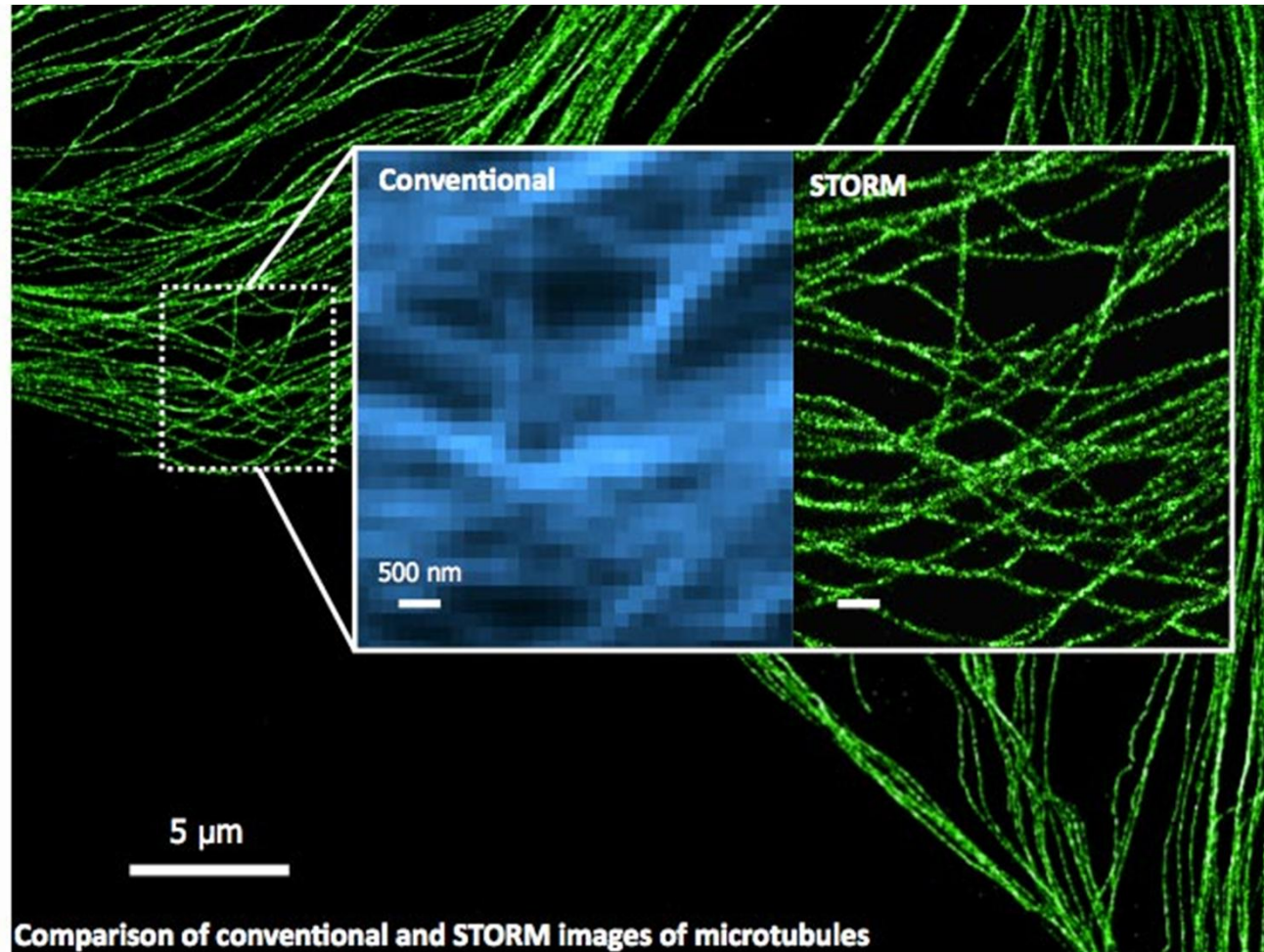


# Comparison of Conventional and STORM Images of Mitochondria in a Mammalian Cell

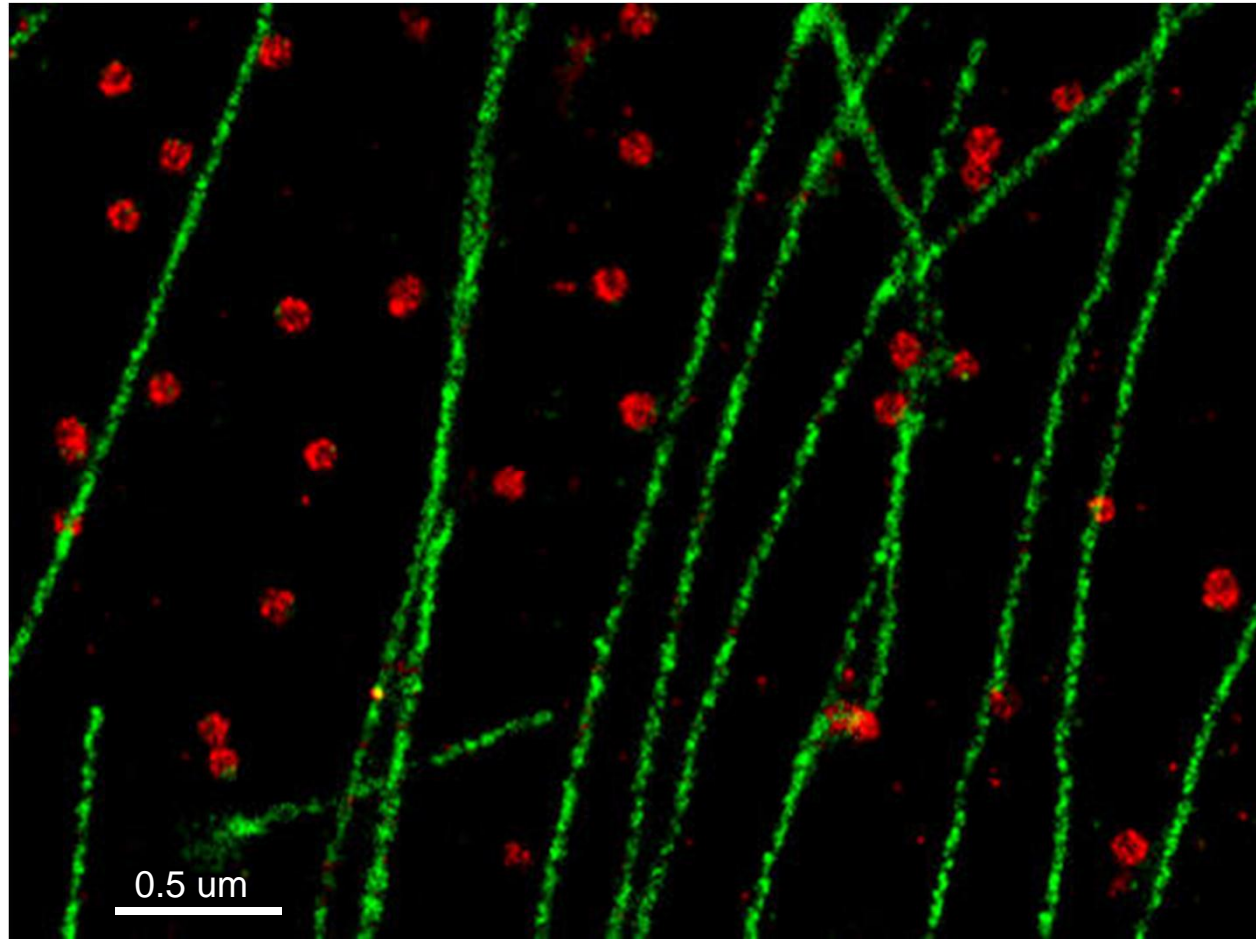




# Comparison of Conventional and STORM Images of microtubules



# Multi-color STORM image of microtubules (green) and clathrin-coated pit (red) in a cell

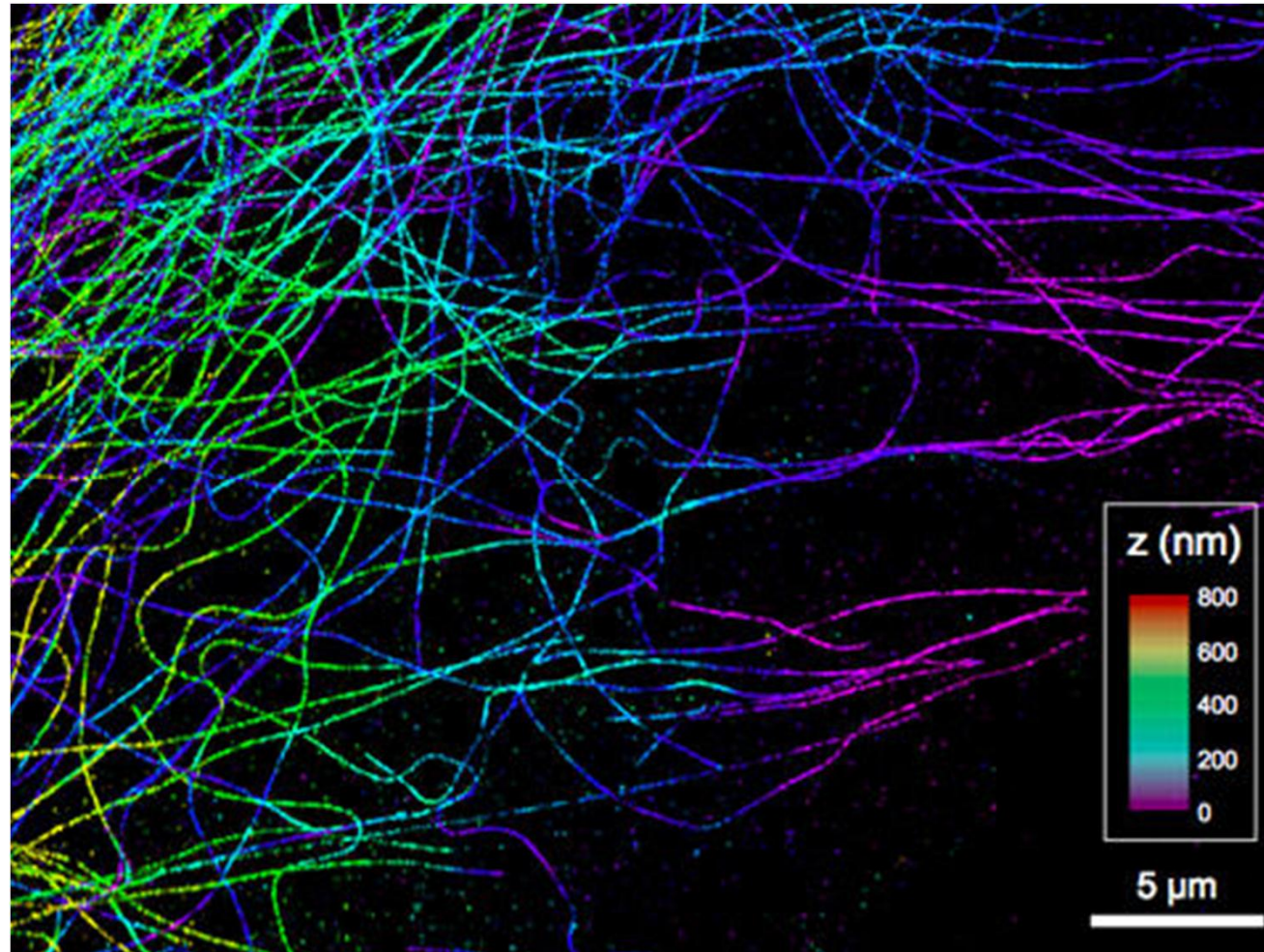


NIKON CORPORATION WEB SITE

Image courtesy of Zhuang Research Group, Department of Chemistry and Chemical Biology, Harvard University, Cambridge, MA.



## 3D STORM Image of Microtubules in a Cell



NIKON CORPORATION WEB SITE

Image courtesy of Zhuang Research Group, Department of Chemistry and Chemical Biology, Harvard University, Cambridge, MA.